

Coronavirus phylogeny based on a geometric approach

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Abstract

A novel coronavirus has been identified as the cause of the outbreak of severe acute respiratory syndrome (SARS). Previous phylogenetic analyses based on sequence alignments show that SARS-CoVs form a new group distantly related to the other three groups of previously characterized coronaviruses. In this paper, a geometric approach based on the Z-curve representation of the whole genome sequence is proposed to analyze the phylogenetic relationships of coronaviruses. The evolutionary distances are obtained through measuring the differences among the three-dimensional Z-curves. The Z-curve is approximately described by its geometric center and the associated three eigenvectors, which indicate the center position and the trend of the Z-curve, respectively. Although some information is lost due to the approximate description of the Z-curve, the phylogenetic tree constructed based on these parameters is consistent with those of previous analyses. The present method has the merits of simplicity and intuitiveness, but it is still in its premature stage. Because the phylogenetic relationships are inferred from the whole genome, instead of some individual genes, the present method represents a new direction of phylogeny study in the post-genome era.

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1. Introduction

The outbreak of atypical pneumonia, referred to as severe acute respiratory syndrome (SARS) was first identified in Guangdong Province, China, and spread to several countries later (Drosten et al., 2003; Ksiazek et al., 2003; Lee et al., 2003; Peiris et al., 2003; Poutanen et al., 2003; Tsang et al., 2003). A novel coronavirus was isolated and found to be the cause of SARS. Although SARS has been under control, some scattering cases infected by SARS-CoVs were reported. No effective drugs are currently available to cure this disease. Gaining insight into the phylogenetic relationships among coronaviruses would be helpful to discover drugs and develop vaccines against the virus.

The SARS-coronavirus is a new member of the order *Nidovirales*, family *Coronaviridae*, and genus *Coronavirus*. They consist of a diverse group of large, enveloped, positive-stranded RNA viruses that cause respiratory and enteric diseases in humans and other animals (Rota et al., 2003). Excluding SARS-CoVs, coronaviruses can be divided into three groups according to serotypes. Group I and group II contain mammalian viruses, while group II coronaviruses contain a hemagglutinin esterase gene homologous to that of Influenza C virus (Lai and Holmes, 2001). Group III contains only avian viruses. Previous work showed that SARS-CoVs are not closely related to any of the previously characterized coronaviruses and form a distinct group (group IV) within the genus *Coronavirus* (Marra et al., 2003; Rota et al., 2003).

An intuitive method is proposed to infer the phylogenetic relationships of coronaviruses in this article. Historically, Cork et al. proposed a three-dimensional

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representation of genomic sequences, called the *W*-curve (Wu et al., 1993). Since then, the *W*-curve has been used to analyze genomic sequences and study the phylogeny of bacteria (Cork, 2003; Cork et al., 2002; Cork and Toguem, 2002). Instead of the sequence alignment, we adopt a geometric method based on the *Z*-curve of the whole genome. The *Z*-curve is a three-dimensional space curve constituting the *unique* representation of a given DNA sequence in the sense that each can be reconstructed given the other (Zhang and Zhang, 1991, 1994). Based on the *Z*-curve method, a coronavirus-specific gene-finding system ZCURVE_CoV has been developed (Chen et al., 2003), and the software is especially suitable for gene recognition in SARS-CoV genomes. The system is further improved by taking the prediction of cleavage sites of viral proteinases in polyproteins into consideration (Gao et al., 2003). Here we use the differences between the three-dimensional space curves as the foundation to derive the phylogeny of coronaviruses. The key problems are what parameters should be used to describe a curve and how to determine evolutionary distances among organisms based on a group of curves. In this paper, we use a series of parameters, such as the geometric center and the covariance matrix to reflect the center position and the distribution pattern of a curve, respectively. The result shows that SARS-CoVs form an independent group, which is consistent with previous analyses.

2. Materials and methods

2.1. Materials

The 24 complete coronavirus genomes used in this paper were downloaded from GenBank, of which 12 are SARS-CoVs and 12 are from other groups of coronaviruses. The name, accession number, abbreviation, and genome length for the 24 genomes are listed in Table 1. According to the existing taxonomic groups, sequences 1–3 belong to group I, and sequences 4–11 are members of group II, while sequence 12 is the only representative of group III. Refer to Table 1 for details.

2.2. The *Z*-curve

The *Z*-curve is a three-dimensional curve that constitutes a unique representation of a given DNA sequence in the sense that each can be uniquely reconstructed given the other (Zhang and Zhang, 1991, 1994). The resulting curve has a zigzag shape, hence the name *Z*-curve. The *Z*-curve is briefly presented as follows. Consider a DNA sequence read from the 5' to the 3'-end with N bases. Beginning from the first base, inspect the sequence one base at a time. In the n th step, where $n = 1, 2, \dots, N$, count the *cumulative* numbers of the bases A, C, G, and T, occurring in the subsequence from the first base to the n th base in the DNA sequence inspected, and denote them by A_n , C_n , G_n , and T_n respec-

Table 1
The accession number, abbreviation, name, and length for each of the 24 coronavirus genomes

No.	Accession	Group	Abbreviation	Genome	Length (nt)
1	NC_002645	I	HCoV-229E	Human coronavirus 229E	27,317
2	NC_002306	I	TGEV	Transmissible gastroenteritis virus	28,586
3	NC_003436	I	PEDV	Porcine epidemic diarrhea virus	28,033
4	U00735	II	BCoV-M	Bovine coronavirus strain Mebus	31,032
5	AF391542	II	BCoV-L	Bovine coronavirus isolate BCoV-LUN	31,028
6	AF220295	II	BCoV-Q	Bovine coronavirus strain Quebec	31,100
7	NC_003045	II	BCoV	Bovine coronavirus	31,028
8	AF208067	II	MHV-M	Murine hepatitis virus strain ML-10	31,233
9	AF201929	II	MHV-2	Murine hepatitis virus strain 2	31,276
10	AF208066	II	MHV-P	Murine hepatitis virus strain Penn 97-1	31,112
11	NC_001846	II	MHV	Murine hepatitis virus	31,357
12	NC_001451	III	IBV	Avian infectious bronchitis virus	27,608
13	AY278488	IV	BJ01	SARS coronavirus BJ01	29,725
14	AY278741	IV	Urbani	SARS coronavirus Urbani	29,727
15	AY278491	IV	HKU-39849	SARS coronavirus HKU-39849	29,742
16	AY278554	IV	CUHK-W1	SARS coronavirus CUHK-W1	29,736
17	AY282752	IV	CUHK-Su10	SARS coronavirus CUHK-Su10	29,736
18	AY283794	IV	SIN2500	SARS coronavirus Sin2500	29,711
19	AY283795	IV	SIN2677	SARS coronavirus Sin2677	29,705
20	AY283796	IV	SIN2679	SARS coronavirus Sin2679	29,711
21	AY283797	IV	SIN2748	SARS coronavirus Sin2748	29,706
22	AY283798	IV	SIN2774	SARS coronavirus Sin2774	29,711
23	AY291451	IV	TW1	SARS coronavirus TW1	29,729
24	NC_004718	IV	TOR2	SARS coronavirus	29,751

tively. The Z-curve consists of a series of nodes P_n , where $n = 1, 2, \dots, N$, whose coordinates are uniquely determined by the Z-transform of DNA sequences (Zhang and Zhang, 1991, 1994)

$$\begin{aligned} x_n &= (A_n + G_n) - (C_n + T_n) \equiv R_n - Y_n, \\ y_n &= (A_n + C_n) - (G_n + T_n) \equiv M_n - K_n, \\ z_n &= (A_n + T_n) - (C_n + G_n) \equiv W_n - S_n, \\ n &= 0, 1, \dots, N, \quad x_n, y_n, z_n \in [-N, N], \end{aligned} \quad (1)$$

where $A_0 = C_0 = G_0 = T_0 = 0$ and $x_0 = y_0 = z_0 = 0$. Here R, Y, M, K, W, and S represent the bases of puRine, pYrimidine, aMino, Keto, Weak hydrogen bonds, and Strong hydrogen bonds, respectively, according to the Recommendation 1984 by the NC-IUB (Cornish-Bowden, 1985). The line that connects the nodes P_0 ($P_0 = 0$), P_1 , P_2, \dots , until P_N one by one sequentially is called the Z-curve for the DNA sequences inspected. The Z-curve defined above is a three-dimensional space curve, having three independent components, i.e., x_n , y_n , and z_n , which display the distributions of bases of R/Y, M/K, and W/S types, respectively, along the sequence. By viewing the Z-curve, some global and local features of the sequence can be detected in a perceivable way. For almost all genome or chromosome sequences, the curves of $z_n \sim n$ are roughly straight lines (Zhang

et al., 2001). For convenience, the curve of $z_n \sim n$ is fitted by a straight line using the least square technique

$$z = kn, \quad (2)$$

where (z, n) is the coordinate of a point on the fitted straight line and k is its slope. Instead of using the curve of $z_n \sim n$, we will use the $z'_n \sim n$ curve hereafter, where

$$z'_n = z_n - kn. \quad (3)$$

2.3. Algorithm

In this paper, we propose a new way to infer evolutionary distances between organisms from the whole genome sequences. As the Z-curve is a unique representation of a genome, it can be used to reflect a genome's characteristics (Fig. 1). For convenience, we use the coordinates (X, Y, Z') rather than (X, Y, Z) . The differences among the Z-curves of these genomes form the basis for constructing the phylogenetic tree. To study the phylogenetic relationships, the process can be separated into three stages. First, the Z-curve of each genome is described by a set of parameters; second, the distance matrix is generated based on the parameters obtained in the first stage; and finally, the phylogenetic tree can be constructed based on the distance matrix.

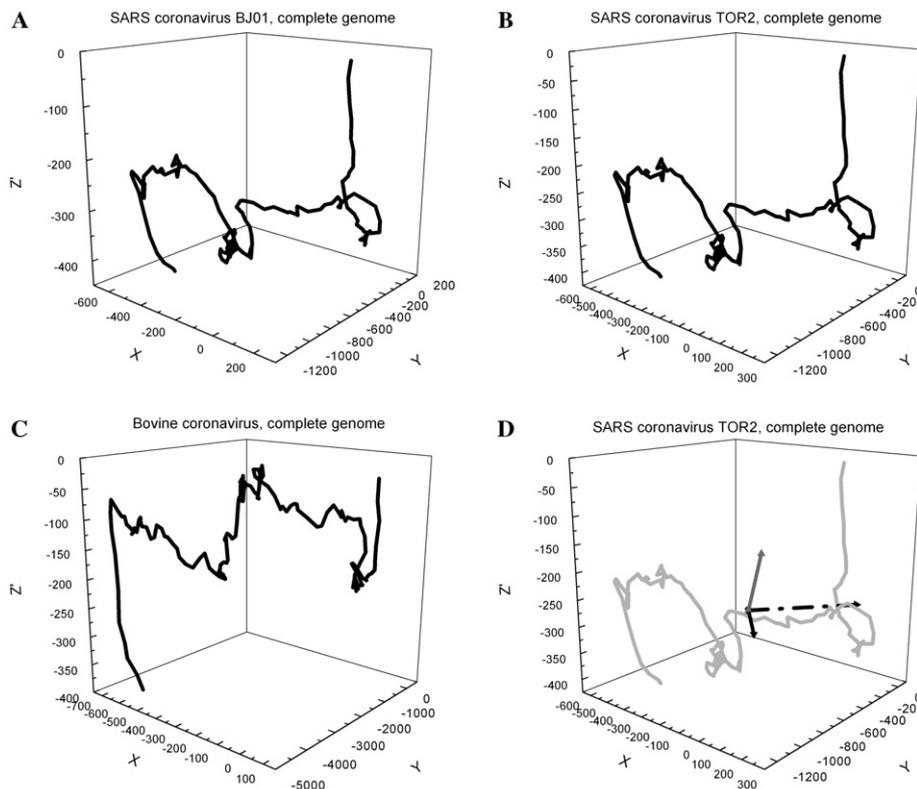


Fig. 1. The three-dimensional Z-curves (x, y, z') for three complete coronavirus genomes. (A–C) The Z-curves of BJ01, TOR2, and BCoV, respectively. It can be clearly seen that the Z-curves of BJ01 and TOR2 are very similar, while the Z-curve of BCoV is significantly different from the former two. This forms the basis of the present method. (D) A sketch of the three eigenvectors for a certain genome (TOR2), which illustrates the relationship between the three eigenvectors and the Z-curve.

Table 2
The geometric center and three eigenvectors of the Z-curve for each of the 24 coronavirus genomes^a

<i>i</i>	Abbreviation	\bar{x}	\bar{y}	\bar{z}'	\mathbf{C}_x^i			\mathbf{C}_y^i			$\mathbf{C}_{z'}^i$		
					$C_{x,x}^i$	$C_{x,y}^i$	$C_{x,z'}^i$	$C_{y,x}^i$	$C_{y,y}^i$	$C_{y,z'}^i$	$C_{z',x}^i$	$C_{z',y}^i$	$C_{z',z'}^i$
1	HCoV-229E	-313.52	-1930.76	22.81	0.80520	-0.16770	-0.56879	0.20016	0.97975	-0.00552	0.55820	-0.10941	0.82246
2	TGEV	-21.86	-1185.34	-87.64	0.95040	-0.06255	-0.30468	0.06443	0.99791	-0.00388	0.30429	-0.01594	0.95245
3	PEDV	-733.04	-1930.98	-194.31	0.90732	-0.36443	-0.20968	0.37249	0.92804	-0.00112	0.19500	-0.07709	0.97777
4	BCoVM	-272.66	-2691.17	-94.48	0.95060	-0.13311	-0.28044	0.13749	0.99049	-0.00411	0.27833	-0.03465	0.95986
5	BCoVL	-268.51	-2658.04	-95.32	0.95068	-0.13276	0.28034	0.13965	0.99019	-0.00464	-0.27697	0.04356	0.95989
6	BCoVQ	-257.34	-2710.49	-90.44	0.95950	-0.13084	-0.24949	0.13399	0.99097	-0.00440	0.24781	-0.02921	0.96837
7	BCoV	-269.55	-2643.97	-95.53	0.97838	-0.13953	0.15268	0.14190	0.98987	-0.00467	-0.15048	0.02624	0.98826
8	MHVM	-129.63	-2295.56	-438.75	0.96108	-0.06279	0.26906	0.06409	0.99794	0.00395	-0.26875	0.01345	0.96312
9	MHV2	-184.66	-2375.26	-428.28	0.98451	-0.07886	0.15662	0.07893	0.99686	0.00582	-0.15659	0.00663	0.98764
10	MHVP	-197.59	-2384.87	-384.67	0.98842	-0.08099	0.12835	0.08133	0.99668	0.00255	-0.12813	0.00792	0.99173
11	MHV	-124.73	-2284.70	-436.33	0.95624	-0.06424	0.28543	0.06586	0.99782	0.00393	-0.28506	0.01504	0.95839
12	IBV	142.03	-1500.55	-289.60	0.72139	0.02288	0.69215	-0.02895	0.99958	-0.00286	-0.69192	-0.01797	0.72175
13	BJ01	-150.58	-627.60	-274.85	0.68326	-0.36116	-0.63460	0.46347	0.88610	-0.00527	0.56422	-0.29052	0.77282
14	Urbani	-152.91	-632.95	-270.13	0.67899	-0.35342	-0.64348	0.45769	0.88910	-0.00538	0.57402	-0.29086	0.76544
15	HKU-39849	-154.55	-622.44	-273.25	0.65621	-0.34854	-0.66926	0.46481	0.88539	-0.00535	0.59442	-0.30757	0.74301
16	CUHK-W1	-150.87	-623.56	-276.66	0.67352	-0.35379	-0.64901	0.46128	0.88724	-0.00495	0.57758	-0.29604	0.76077
17	CUHK-Su10	-150.06	-626.75	-278.13	0.67051	-0.34988	-0.65422	0.45881	0.88852	-0.00496	0.58302	-0.29684	0.75629
18	SIN2500	-148.99	-627.23	-277.78	0.67073	-0.35031	-0.65377	0.45932	0.88826	-0.00472	0.58237	-0.29712	0.75668
19	SIN2677	-148.91	-629.56	-278.38	0.66552	-0.34716	-0.66073	0.45863	0.88862	-0.00494	0.58885	-0.29974	0.75061
20	SIN2679	-148.22	-627.18	-277.99	0.66644	-0.34686	-0.65995	0.45800	0.88894	-0.00471	0.58829	-0.29912	0.75129
21	SIN2748	-149.34	-626.83	-277.39	0.66698	-0.34948	-0.65803	0.46041	0.88769	-0.00479	0.58580	-0.29977	0.75298
22	SIN2774	-148.27	-627.13	-277.97	0.66611	-0.34675	-0.66035	0.45810	0.88889	-0.00467	0.58859	-0.29939	0.75095
23	TW1	-152.93	-632.21	-272.52	0.66820	-0.34900	-0.65705	0.45918	0.88833	-0.00488	0.58538	-0.29844	0.75383
24	TOR2	-152.69	-630.34	-271.00	0.67042	-0.34999	-0.65425	0.45892	0.88846	-0.00502	0.58303	-0.29688	0.75627

^a Refer to the text for detailed explanation about the meaning of the mathematical symbols used in this table.

(i) *The parameters of the Z-curve for each genome.* Based on the Z-curve, any genome can be represented by a three-dimensional space curve composed of N nodes corresponding to every base position denoted by x_n, y_n, z'_n where $n = 1, 2, \dots, N$ (Figs. 1A–C). To describe its characteristics, we calculate the following parameters. The first is the geometric center of all the n nodes

$$\bar{x} = \frac{1}{N} \sum_{n=1}^N x_n, \quad \bar{y} = \frac{1}{N} \sum_{n=1}^N y_n, \quad \bar{z}' = \frac{1}{N} \sum_{n=1}^N z'_n. \quad (4)$$

Consequently, we can obtain $(\bar{x}, \bar{y}, \bar{z}')$ for each genome. Refer to Table 2 for details.

Then, the covariance matrix which describes the global distribution pattern of the three-dimensional space curve is calculated as follows:

$$\Sigma = \begin{pmatrix} \sigma_{xx} & \sigma_{xy} & \sigma_{xz'} \\ \sigma_{yx} & \sigma_{yy} & \sigma_{yz'} \\ \sigma_{z'x} & \sigma_{z'y} & \sigma_{z'z'} \end{pmatrix} = (\sigma_{pq}), \quad p, q = x, y, z', \quad (5)$$

where

$$\sigma_{pq} = \frac{1}{N-1} \sum_{n=1}^N (p_n - \bar{p})(q_n - \bar{q}), \quad (6)$$

where $p, q = x, y, z'$.

Obviously, the matrix is a real symmetric 3×3 one. Using a 3×3 matrix to represent a three-dimensional Z-curve is a very rough approximation, resulting in information loss considerably. However, the advantage is that this approximation makes it possible to compare genomes with different lengths. It is seen that a 3×3 covariance matrix is uniquely derived based on Eq. (6) for each given genome regardless of its length. From a geometrical point of view, the distribution pattern can be reduced to a three-dimensional ellipsoid approximately. Each direction of the main axis of the ellipsoid can be denoted by an eigenvector and its length should be proportional to the square root of its associated eigenvalue. The eigenvectors and their associated eigenvalues are defined as follows:

$$\Sigma \mathbf{C}_k = \lambda_k \mathbf{C}_k, \quad \mathbf{C}_k = (C_{k,1}, C_{k,2}, C_{k,3})^T, \quad k = 1, 2, 3. \quad (7)$$

Corresponding to each eigenvalue λ_k , there's an eigenvector \mathbf{C}_k . Corresponding to $\lambda_1 < \lambda_2 < \lambda_3$, the three eigenvectors are denoted by $\mathbf{C}_1, \mathbf{C}_2, \mathbf{C}_3$, respectively. It's easy to obtain the eigenvalues and associated normalized eigenvectors using the Jacobi algorithm. The geometric center and three eigenvectors for each of the 24 genomes are obtained in the same way. Refer to Table 2 for details about the parameters.

(ii) *The distance matrix derived from the above parameters.* In this paper, the Euclid distance is used to reflect the diversity between two points

$$d_{ij} = \sqrt{(\bar{x}_i - \bar{x}_j)^2 + (\bar{y}_i - \bar{y}_j)^2 + (\bar{z}'_i - \bar{z}'_j)^2}, \quad i, j = 1, 2, \dots, M, \quad (8)$$

where d_{ij} denotes the distance between the geometric centers of the i th and the j th genomes, and M is the total number of all genomes ($M = 24$, here). Then we obtain a real $M \times M$ symmetric matrix whose elements are d_{ij} .

To reflect the differences between the trends of every two three-dimensional curves, the angles between the corresponding eigenvectors of every two genomes are used. The three-dimensional vectors are denoted as follows:

$$\mathbf{C}_k^i = (C_{k,x}^i, C_{k,y}^i, C_{k,z'}^i)^T, \quad i = 1, 2, \dots, M, \quad k = 1, 2, 3, \quad (9)$$

where \mathbf{C}_k^i is the k th vector of the i th genome. Each genome has three such eigenvectors. According to the projections on the three axes, the vectors can be divided into three groups. The three groups of vectors are represented with arrows of different styles (refer to Fig. 2A). Obviously they can be separated apart depending on their space distribution. The dark group (X group) has the greatest projections on the x -axis, while the vectors represented with dot (Y group) and grey (Z' group) arrows have the greatest projections on the y -axis and the z' -axis, respectively. For each genome, the three vectors can be divided into three groups, i.e., each genome has three vectors belonging to three groups, respectively.

The three groups of eigenvectors are obtained, and denoted by $\mathbf{C}_x^i, \mathbf{C}_y^i, \mathbf{C}_{z'}^i$, respectively (see Table 2). The cosine between any two vectors in a certain group can be computed as follows:

$$\cos \theta_{ij}^k = \frac{\mathbf{C}_k^i \cdot \mathbf{C}_k^j}{|\mathbf{C}_k^i| \cdot |\mathbf{C}_k^j|}, \quad i, j = 1, 2, \dots, M, \quad k = x, y, z'. \quad (10)$$

Repeating this procedure for all the three groups, we obtain three real $M \times M$ symmetric matrices. These matrices are then translated into angles, whose elements are as follows:

$$\theta_{ij}^k = \arccos(\cos \theta_{ij}^k), \quad i, j = 1, 2, \dots, M, \quad k = x, y, z'. \quad (11)$$

The sum of θ_{ij}^k over k for given i, j can be used to reflect the trend information of the eigenvectors involved

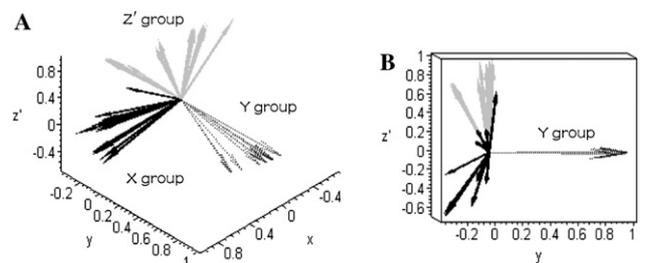


Fig. 2. The three groups of eigenvectors (denoted with different arrows). The vectors in the X, Y, and Z' groups are denoted by dark, dot, and grey arrows, respectively. (A and B) The eigenvectors of the 24 genomes observed from different directions. It can be seen from (A) that the three groups can be separated according to their three-dimensional space distribution. (B) The vectors in Y group of the 24 genomes are coplanar and they are almost in the x - y plane.

$$\Theta_{ij} = \theta_{ij}^x + \theta_{ij}^y + \theta_{ij}^z, \quad i, j = 1, 2, \dots, M. \quad (12)$$

Consequently, two sets of parameters are obtained. The first reflects the difference of center positions represented by the Euclid distance between the geometric centers. The second indicates the difference of the trends of the Z -curves represented by the related eigenvectors. The overall distance D_{ij} between the species i and j is defined by

$$D_{ij} = d_{ij} \times \Theta_{ij}, \quad i, j = 1, 2, \dots, M. \quad (13)$$

(iii) *Clustering*. Accordingly, a real symmetric $M \times M$ matrix D_{ij} is obtained and used to reflect the evolutionary distance between the species i and j . The clustering tree is constructed using the UPGMA method in PHYLIP package (<http://evolution.genetics.washington.edu/phylip.html>). The final phylogenetic tree is drawn using the DRAWGRAM program in the PHYLIP package. The branch lengths are not scaled according to the distances and only the topology of the tree is concerned.

3. Results and discussion

3.1. The three-dimensional Z -curve for a complete genome

As mentioned above, one of the advantages of the Z -curve is its intuitiveness. The feature of a genome can be viewed intuitively regardless of how long the genome is. Therefore, global and local compositional features of a genome can be grasped quickly in a perceivable form (Zhang et al., 2003). To give an intuitive comprehension of the difference among the three-dimensional curves, we take SARS-CoV strains TOR2, BJ01, and BCoV as examples. TOR2 and BJ01 are SARS-CoVs and BCoV belongs to another group of coronaviruses. From the coordinates and the trends in Figs. 1A–C, we can see that the Z -curves of TOR2 and BJ01 are almost the same while that of BCoV is significantly different from both of them, indicating that the former two have close phylogenetic relationship, whereas the relationships between the former two and the latter are more distant. Similarity of related Z -curves implies close evolutionary relationship of the organisms involved (Zhang et al., 2003) and vice versa. This constitutes the basis of the current algorithm.

The Z -curve is approximately described by the geometric center and eigenvectors, which indicate its center position and the trends, respectively (Fig. 1D). In Fig. 1D the three arrows represent the three eigenvectors, and the point from which they start is the geometric center. The three eigenvectors of a certain genome can be divided into three groups according to their relationships with the axes (refer to Fig. 2). The trends of Z -curves carry a part of the information used to construct the phylogenetic tree, and some interesting results can be

revealed by this figure. It can be seen from Fig. 2B that the vectors in the Y group, which have the greatest projections on the positive y -axis, are coplanar perfectly. They are almost in the x - y plane. As can be seen from the plot, the 24 vectors are almost superposed with each other as a single vector. The phenomenon can also be seen from the data in Table 2. All of the absolute value of $C_{y,z}^i$ ($i = 1, 2, \dots, M$) are smaller than 0.0059. That is to say, they all have very small projections on the z' -axis and are constrained into the x - y plane. The vectors in the X group and Z' group (represented with black and grey arrows, respectively, in Fig. 2B) are also coplanar in the x - z' plane, though their coplanarity is not as good as that of the Y group.

3.2. Phylogenetic tree of coronaviruses

As mentioned above, there are three groups of coronaviruses. Group I includes HCoV-229E, TGEV, and PEDV and group II contains BCoV, BCoV-L, BCoV-M, BCoV-Q, MHV, MHV2, MHV-M, MHV-P, etc. All the viruses in these two groups are mammalian viruses. Group III contains only avian viruses, of which only the genome of IBV has been completely sequenced. Many researchers have analyzed the phylogenetic relationships among coronavirus genomes based on the 3C-like proteinase, polymerase, the structural proteins S, E, M, and N, respectively (Marra et al., 2003; Rota et al., 2003). Their results indicated that SARS-CoVs are not closely related to any of the previously characterized coronaviruses and form a distinct group (group IV) within the genus *Coronavirus* (Marra et al., 2003; Rota et al., 2003). As shown in Fig. 3, four groups of coronaviruses can be seen from the phylogram. The SARS-CoVs appear to cluster together and form a separate branch, which can be distinguished easily from other three groups of coronaviruses. IBV, belonging to group III, is situated at an independent branch, whereas the TGEV, PEDV, and HCoV-229E, which belong to group I, tend to cluster together. In another branch, the group II coronaviruses, including BCoV, BCoV-L, BCoV-M, BCoV-Q, MHV, MHV2, MHV-M, and MHV-P tend to cluster together. First, group I and group II, which are all mammalian viruses, cluster together forming a bigger group. Second, this group joins group III, which contains only avian viruses, to form a much bigger group. Finally, SARS-CoVs join them and result in the phylogenetic tree shown in Fig. 3. The resulting monophyletic clusters agree perfectly with the established taxonomic groups. To validate the current method, a set of random sequences were used as a control. We generated 100 random sequences meeting the requirements in the method. Each time a phylogenetic analysis was done using 25 sequences including one random sequence and the 24 genomes. Consequently, 100 phylogenetic trees were obtained. Ninety-eight out of

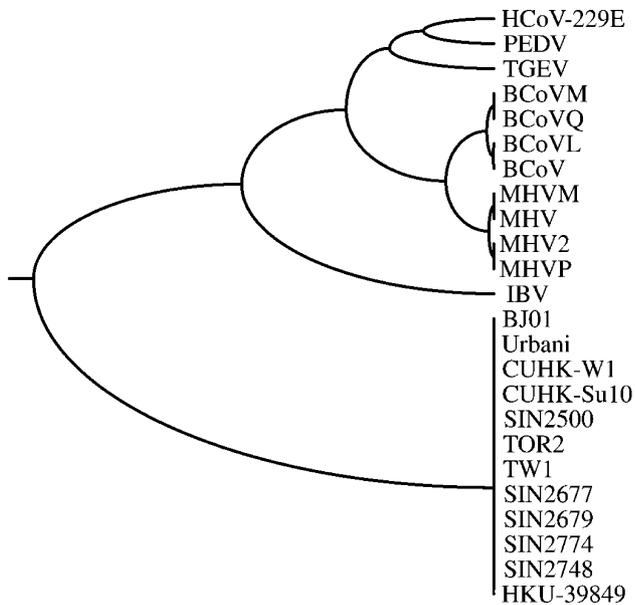


Fig. 3. The phylogenetic tree constructed with the current method. The result shows that four groups exist in the genus *Coronavirus*. Note that group I (HCoV-229E, TGEV, and PEDV) and group II (BCoVM, BCoV, BCoVQ, BCoV, MHVM, MHV2, MHVP, and MHV) cluster together forming a bigger group firstly. Second, this group joins group III (IBV) to form a much bigger group. Finally, SARS-CoVs join them and result in the phylogenetic tree shown here. Also note that the resulting monophyletic clusters agree perfectly with the established taxonomic groups.

the 100 trees showed that the random sequence formed a distinct group without disturbing the other four groups. Only two of the random sequences disturbed the four groups, suggesting that the current method is solid with respect to the situation that a random sequence is added.

3.3. Comparison with the results of previous analyses

Almost all of the previous analyses revealed that SARS-CoVs form a distinct group different from the other three groups of coronaviruses. However, the question that how SARS-CoVs emerged suddenly still remains open. Rota et al. and Marra et al. performed phylogenetic analysis based on sequence alignments using different genes. The results indicated that SARS-CoVs belong to a new group but the original group that SARS-CoVs were derived from could not be determined (Rota et al., 2003). The detection of SARS-CoV-like viruses in Himalayan palm civets and other small animals in live retail market indicates a route of interspecies transmission, although the natural reservoir is unknown. Virus infection was also detected in humans working at the same market. All the animal isolates retain a special 29-nucleotide fragment, which is not found in most human isolates (Guan et al., 2003). Stavrinides and Guttman made phylogenetic analysis on the SARS virus replicase, surface spike, matrix, and nucleocapsid proteins. The results support a mammalian-like origin

for the replicase protein, an avian-like origin for the matrix and nucleocapsid proteins, and a mammalian–avian mosaic origin for the host-determining spike protein. They proposed that a recombination event between mammalian-like and avian-like parent viruses within the S gene might have taken place (Stavrinides and Guttman, 2004). However, the phylogenetic inference based on genome contents tends to locate the recombinant outside of related genomes, such as seen in Fig. 3. Therefore, we emphasize that it is very unlikely to trace back the evolutionary history such as the recombination event using the method presented.

The present method reflects the global characters of genomes because the whole genome is taken into consideration. The phylogenetic tree (Fig. 3) reveals that the SARS-CoVs have undergone an independent evolution path after the divergence from the other coronaviruses. As can be seen from Fig. 3, the distance between the SARS-CoVs and all the others is the greatest. We supposed that the precursor of SARS-CoV may have existed in some hosts and developed separately for many years. Grigoriev found that the mutational patterns in SARS-CoV genome were strikingly different from the other coronaviruses in terms of mutation rates (Grigoriev, 2004). Phylogenetic analysis based on codon usage pattern suggested that SARS-CoV was diverged far from all the three known groups of coronavirus (Gu et al., 2004). The overall level of similarity between SARS-CoVs and the other coronaviruses is low (Rota et al., 2003). We suppose that this is due to different evolution paths. The isolation of SARS-CoV-like virus in Himalayan palm civets indicates a route of interspecies transmission. We hypothesize that some events such as the nucleotide deletion or mutation in some important genes of the precursor may have resulted in the change of host range.

3.4. Merits of the current method

Due to the lack of morphological features and frequent gene exchanges, it is highly valuable to develop methods of molecular phylogeny for viruses. Now phylogenetic analysis based on sequence alignments is well developed. Sequence alignments are always based on some special genes or some conserved fragments (Saitou, 1996). Such analysis can be done at both the amino acid level and the nucleotide level. To overcome the biases caused by individual genes or genome segments, it is valuable to develop methods of molecular phylogenetic analysis based on whole genome sequences. Being different from the sequence alignment method, the current method is a geometric approach which is based on measuring the differences of Z-curves of whole genomes, including coding and non-coding sequences. There is no need to search for similar sequences. Probably, the most remarkable advantages of the present method is its simplicity and intuitiveness.

The increasing availability of complete genomes has cast doubt instead of adding details to the phylogenetic tree (Qi et al., 2004). Phylogenetic analysis based on sequence alignments is usually done on the most conservative part of a gene. These fragments are usually coding sequences, especially the sequences coding for catalytic sites or the core of proteins, because they tend to be more evolutionarily conserved. It was said by a virologist that people could not simply assume that a virus can be represented by its polymerase (<http://www.ncbi.nlm.nih.gov/ICTV/>). A virus must be viewed as a whole. Non-coding sequences also play an important role in the virus, so do the less conserved genes. In addition, analyses based on different genes may lead to different results. Consequently, by using complete genomes one can avoid choosing which genes to be aligned. Therefore methods that are based on the whole genome are likely to be more objective. Recently, a *k*-string composition approach was proposed to analyze prokaryote phylogeny based on the whole proteome and satisfactory results were obtained (Qi et al., 2004); however, such analysis must rely on the annotation information. In contrast, the complete genome sequence is the only input of the current method; neither the annotation information, nor any adjustable parameters are needed. It is noteworthy that the current method is performed automatically without any human intervention.

The *Z*-curve, which serves as the foundation of the present method is a powerful tool to study the complete genome sequence. The *Z*-curve contains all the information that the corresponding DNA sequence carries. Many characteristics of a genome with biological meaning can be observed from the corresponding *Z*-curve, such as the replication origins and genomic islands for some bacterial and archaeal genomes (Zhang et al., 2003). We can inspect a genome in an intuitive way regardless of the gene content and gene order, even though the sequences are of different lengths. If the *Z*-curves of two species show similar pattern even though the genomes have different lengths, one may infer that they are evolutionarily close organisms, and vice versa. In this paper, we use the geometric center and the eigenvectors to describe the pattern approximately. Although this is only a rough approximation, it represents just an attempt to apply the *Z*-curve method to the phylogenetic analysis and the results obtained agree well with previous analyses.

3.5. Limitations of the current method

This method is aimed to analyze the phylogeny of the genomes which have close phylogenetic relationships. Phylogenetics analysis is based on the differences among the three-dimensional *Z*-curves. In this paper, the 24 genomes under study all belong to the same genus *Coro-*

navirus. Additionally, the differences of length among genomes are not very large. If the genomes under study have much farther phylogenetic relationships, and the differences in length are considerably large, the present method may not work. Consequently, cautions must be taken when using the present method to study the phylogeny of organisms with far evolutionary distances. In addition, unlike the estimation based on comparison of orthologous genes, the *Z*-curve approach is also sensitive to genome rearrangements: a single large-scale inversion can change the form of *Z*-curve drastically. Therefore, the method presented here is considerably limited in the cases of genome rearrangements. In addition, as mentioned above, the three-dimensional *Z*-curve is approximately depicted by a few parameters, such as the geometric center and the associated three eigenvectors. Consequently, information contained in the *Z*-curve is lost considerably in so doing. It is reasonable to suppose that the more information is extracted from the *Z*-curve, the more accurate result can be gained. Therefore, the current method can be improved if new and more effective algorithms are proposed to extract information contained in the *Z*-curves. In summary, although the present method has some advantages, it is still in its premature stage. The method may not be applied to some general cases, therefore the applications of it are considerably limited at present.

4. Conclusion

A geometric approach to infer phylogenetic relationships based on the *Z*-curves of complete genomes is proposed in this article. Phylogenetic analysis of the 24 coronaviruses shows that SARS-CoVs belong to a new cluster, named group IV, and this result is consistent with those of previous analyses. The method has much room to be improved because of the possibility to extract information from the whole genome, instead of some individual genes. Although having some limitations, the current whole-genome-based geometric approach represents a new direction to infer phylogenetic relationships of organisms in the post-genome era. However, the method is still in its premature stage and its applications are considerably limited at present.

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References

- Chen, L.L., Ou, H.Y., Zhang, R., Zhang, C.-T., 2003. ZCURVE - CoV: a new system to recognize protein coding genes in coronavirus genomes, and its applications in analyzing SARS-CoV genomes. *Biochem. Biophys. Res. Commun.* 307, 382–388.
- Cork, D.J., 2003. Achieving consensus of long genomic sequences with the W-curve. In: Lapointe, F., McMorris, F.R., Janowitz, M. (Eds.), *Bioconsensus, DIMACS Series in Discrete Mathematics and Theoretical Computer Science*, vol. 61. American Mathematical Society, Providence, RI, pp. 123–134.
- Cork, D.J., Hutch, T.B., Marland, E., Zmuda, J., 2002. Achieving congruency of phylogenetic trees generated by W-curves of genomic sequences. In: Valafar, F. (Ed.), *Techniques in Bioinformatics and Medical Informatics*. Ann. N. Y. Acad. Sci. 980, 23–31.
- Cork, D.J., Toguem, A., 2002. Using fuzzy logic to confirm the integrity of a pattern recognition algorithm for long genomic sequences: the W-curve of genomic sequences. In: Valafar, F. (Ed.), *Techniques in Bioinformatics and Medical Informatics*. Ann. N. Y. Acad. Sci. 980, 32–40.
- Cornish-Bowden, A., 1985. Nomenclature for incompletely specified bases in nucleic acid sequences: recommendation 1984. *Nucleic Acids Res.* 13, 3021–3030.
- Drosten, C., Gunther, S., Preiser, W., van der Werf, S., Brodt, H.R., Becker, S., Rabenau, H., Panning, M., Kolesnikova, L., Fouchier, R.A., Berger, A., Burguiere, A.M., Cinatl, J., Eickmann, M., Escriviou, N., Grywna, K., Kramme, S., Manuguerra, J.C., Muller, S., Rickerts, V., Sturmer, M., Vieth, S., Klenk, H.D., Osterhaus, A.D., Schmitz, H., Doerr, H.W., 2003. Identification of a novel coronavirus in patients with severe acute respiratory syndrome. *N. Engl. J. Med.* 348, 1967–1976.
- Gao, F., Ou, H.Y., Chen, L.L., Zheng, W.X., Zhang, C.-T., 2003. Prediction of proteinase cleavage sites in polyproteins of coronaviruses and its applications in analyzing SARS-CoV genomes. *FEBS Lett.* 553, 451–456.
- Grigoriev, A., 2004. Mutational patterns correlate with genome organization in SARS and other coronaviruses. *Trends Genet.* 20, 131–135.
- Gu, W., Zhou, T., Ma, J., Sun, X., Lu, Z., 2004. Analysis of synonymous codon usage in SARS coronavirus and other viruses in the Nidovirales. *Virus Res.* 101, 155–161.
- Guan, Y., Zheng, B.J., He, Y.Q., Liu, X.L., Zhuang, Z.X., Cheung, C.L., Luo, S.W., Li, P.H., Zhang, L.J., Guan, Y.J., Butt, K.M., Wong, K.L., Chan, K.W., Lim, W., Shortridge, K.F., Yuen, K.Y., Peiris, J.S., Poon, L.L., 2003. Isolation and characterization of viruses related to the SARS coronavirus from animals in southern China. *Science* 302, 276–278.
- Ksiazek, T.G., Erdman, D., Goldsmith, C.S., Zaki, S.R., Peret, T., Emery, S., Tong, S., Urbani, C., Comer, J.A., Lim, W., Rollin, P.E., Dowell, S.F., Ling, A.-E., Humphrey, C.D., Shieh, W.-J., Guarner, J., Paddock, C.D., Rota, P., Fields, B., DeRisi, J., Yang, J.-Y., Cox, N., Hughes, J.M., LeDuc, J.W., Bellini, W.J., Anderson, L.J., the SARS Working Group, 2003. A novel coronavirus associated with severe acute respiratory syndrome. *N. Engl. J. Med.* 348, 1953–1966.
- Lai, M.M.C., Holmes, K.V., 2001. *Coronaviridae: the viruses and their replication*. In: Knipe, D.M., Howley, P.M. (Eds.), *Fields Virology*, fourth ed. Lippincott Williams and Wilkins, New York (Chapter 35).
- Lee, N., Hui, D., Wu, A., Chan, P., Cameron, P., Joynt, G.M., Ahuja, A., Yung, M.Y., Leung, C.B., To, K.F., Lui, S.F., Szeto, C.C., Chung, S., Sung, J.J., 2003. A major outbreak of severe acute respiratory syndrome in Hong Kong. *N. Engl. J. Med.* 348, 1986–1994.
- Marra, M.A., Jones, S.J., Astell, C.R., Holt, R.A., Brooks-Wilson, A., Butterfield, Y.S., Khattra, J., Asano, J.K., Barber, S.A., Chan, S.Y., Cloutier, A., Coughlin, S.M., Freeman, D., Girn, N., Griffith, O.L., Leach, S.R., Mayo, M., McDonald, H., Montgomery, S.B., Pandoh, P.K., Petrescu, A.S., Robertson, A.G., Schein, J.E., Siddiqui, A., Smailus, D.E., Stott, J.M., Yang, G.S., Plummer, F., Andonov, A., Artsob, H., Bastien, N., Bernard, K., Booth, T.F., Bowness, D., Czub, M., Drebot, M., Fernando, L., Flick, R., Garbutt, M., Gray, M., Grolla, A., Jones, S., Feldmann, H., Meyers, A., Kabani, A., Li, Y., Normand, S., Stroher, U., Tipples, G.A., Tyler, S., Vogrig, R., Ward, D., Watson, B., Brunham, R.C., Krajden, M., Petric, M., Skowronski, D.M., Upton, C., Roper, R.L., 2003. The genome sequence of the SARS-associated coronavirus. *Science* 300, 1399–1404.
- Peiris, J.S., Lai, S.T., Poon, L.L., Guan, Y., Yam, L.Y., Lim, W., Nicholls, J., Yee, W.K., Yan, W.W., Cheung, M.T., Cheng, V.C., Chan, K.H., Tsang, D.N., Yung, R.W., Ng, T.K., Yuen, K.Y.; SARS study group, 2003. Coronavirus as a possible cause of severe acute respiratory syndrome. *Lancet* 361, 1319–1325.
- Poutanen, S.M., Low, D.E., Henry, B., Finkelstein, S., Rose, D., Green, K., Tellier, R., Draker, R., Adachi, D., Ayers, M., Chan, A.K., Skowronski, D.M., Salit, I., Simor, A.E., Slutsky, A.S., Doyle, P.W., Krajden, M., Petric, M., Brunham, R.C., McGeer, A.J., the National Microbiology Laboratory, Canada, and the Canadian Severe Acute Respiratory Syndrome Study Team, 2003. Identification of severe acute respiratory syndrome in Canada. *N. Engl. J. Med.* 348, 1995–2005.
- Qi, J., Wang, B., Hao, B.L., 2004. Whole proteome prokaryote phylogeny without sequence alignment: a K-string composition approach. *J. Mol. Evol.* 58, 1–11.
- Rota, P.A., Oberste, M.S., Monroe, S.S., Nix, W.A., Campagnoli, R., Icenogle, J.P., Penaranda, S., Bankamp, B., Maher, K., Chen, M.H., Tong, S., Tamin, A., Lowe, L., Frace, M., DeRisi, J.L., Chen, Q., Wang, D., Erdman, D.D., Peret, T.C., Burns, C., Ksiazek, T.G., Rollin, P.E., Sanchez, A., Liffick, S., Holloway, B., Limor, J., McCaustland, K., Olsen-Rasmussen, M., Fouchier, R., Gunther, S., Osterhaus, A.D., Drosten, C., Pallansch, M.A., Anderson, L.J., Bellini, W.J., 2003. Characterization of a novel coronavirus associated with severe acute respiratory syndrome. *Science* 300, 1394–1399.
- Saitou, N., 1996. Reconstruction of genes trees from sequence data. *Methods Enzymol.* 266, 427–449.
- Stavrinos, J., Guttman, D.S., 2004. Mosaic evolution of the severe acute respiratory syndrome coronavirus. *J. Virol.* 78, 76–82.
- Tsang, K.W., Ho, P.L., Ooi, G.C., Yee, W.K., Wang, T., Chan-Yeung, M., Lam, W.K., Seto, W.H., Yam, L.Y., Cheung, T.M., Wong, P.C., Lam, B., Ip, M.S., Chan, J., Yuen, K.Y., Lai, K.N., 2003. A cluster of cases of severe acute respiratory syndrome in Hong Kong. *N. Engl. J. Med.* 348, 1977–1985.
- Wu, D., Roberge, J., Cork, D.J., Nguyen, B.J., Grace, T., 1993. Computer visualization of long genomic sequences. *Proc. IEEE Visualization* 93, 308–315.
- Zhang, C.-T., Wang, J., Zhang, R., 2001. A novel method to calculate the G + C content of genomic DNA sequences. *J. Biomol. Struct. Dyn.* 19 (2), 333–341.
- Zhang, C.-T., Zhang, R., 1991. Analysis of distribution of bases in the coding sequences by a diagrammatic technique. *Nucleic Acids Res.* 19, 6313–6317.
- Zhang, R., Zhang, C.-T., 1994. Z-curves, an intuitive tool for visualizing and analyzing DNA sequences. *J. Biomol. Struct. Dyn.* 11, 767–782.
- Zhang, C.-T., Zhang, R., Ou, H.Y., 2003. The Z-curve database: a graphic representation of genome sequences. *Bioinformatics* 19, 593–599.