297

SARS Coronavirus Anti-Infectives

Tommy R. Tong*

Department of Pathology, Princess Margaret Hospital, Hong Kong

Received: May 15, 2005; Accepted: September 18, 2006; Revised: September 19, 2006

Abstract: Severe acute respiratory syndrome (SARS) emerged in late 2002 and was controlled in July 2003 by public health measures. Its causative agent, SARS coronavirus (SARS-CoV) jumped from an animal reservoir to humans and has the potential to re-emerge. Following the sequencing of the genetic code and the deciphering of some of the functions of its proteins, including the cellular receptors and host proteins that participate in the life cycle of the virus, promising lead drugs and new uses of old drugs have been discovered. Patent applications for cathepsin L inhibitors have taken new relevance because of the role of cathepsin L in the entry of SARS-CoV into host cells. Likewise, patent applications for SARS-CoV protease inhibitors and interferon and mismatched dsRNA also need to be watched for potential application in treatment and prevention of SARS-CoV. Here, we review the recent advances and inventions that target SARS-CoV infection in humans.

Keywords: Severe acute respiratory syndrome, SARS, SARS coronavirus, SARS-CoV, anti-infective, anti-viral, main protease, 3CLpro, polymerase, helicase, interferon, interferon-inducer, antibody.

INTRODUCTION

SARS [1-5] is a viral pneumonia with 10% fatality rate caused by a previously unknown coronavirus (CoV) that crossed and adapted to humans from animal reservoirs [6,7]. The disease emerged in late 2002 and had spread to 29 countries within a few weeks, infecting ~8,000 people and causing some 800 fatalities. International cooperation resulted in the dramatic prevention of a potentially catastrophic pandemic [8]. In the 3 years since the epidemic, the molecular evolution [9] of the virus has been worked out and additional novel coronaviruses identified in humans and animals, greatly increasing our knowledge of the Coronaviridae.

The SARS-CoV genome (Fig. 1) is among the largest in the world of RNA viruses (27-31 kb). It is single-stranded, sense (+), capped and methylated at the 5' end, and polyadenylated at the 3' end. SARS-CoV genome has 14 predicted open reading frames (ORF) encoding 28 proteins [10-12]. Through a putative recombination event with an unidentified virus, SARS-CoV acquired a receptor-binding domain (RBD) that is specific for the non-catalytic region of human angiotensin-converting enzyme 2 (ACE2). It also binds civet ACE2 with avidity [13]. After attaching to ACE2, a necessary and sufficient cellular receptor, SARS-CoV spike undergoes conformational change that leads to fusion of its lipid envelope with the cell membrane. The nucleocapsid enters the cytosolic compartment, where cellular translational machinery begins without delay to produce viral replicase enzymes that self-assemble after auto-proteolytic cleavage of the ORF1 gene product. Polyprotein (pp) 1a is translated from ORF1a. Pp1ab is encoded by an overlapping ORF1a and ORF1b, and is translated by a -1 ribosomal frameshift mechanism. Other ORFs and structural proteins are translated from a nested set of 3'-co-terminal subgenomic mRNAs. Spike (S), envelope (E), and membrane (M) proteins are targeted to intracellular membranes between the endoplasmic reticulum and Golgi apparatus. Replicated viral genomic RNA associate with nucleocapsid (N) proteins, which interact with M, triggering viral assembly. This is followed by budding into vesicles, which traffic to the cell surface, where mature virions are released.

CLASSES OF SARS-COV ANTI-INFECTIVES

More than 30 successful drugs against different viruses are now available, abolishing the notion that viral illnesses cannot be treated specifically. Viral entry, transcription, replication, maturation, and cellular processes usurped by viruses represent possible therapeutic targets. The subject was recently reviewed by De Clercq [14].

However, the challenge now is to be able to respond fast enough to emerging viral diseases as well as to reduce the cost of producing drugs that may only be useful in a small number of subjects - a case of drug therapy competing with public health measures. This quandary was illustrated by the "Katrina-like" emergence and subsidence of SARS-CoV [15]. Table 1 summarizes the various classes of SAR-CoV anti-infectives discussed below.

I. VIRAL ENTRY INHIBITORS

Prevention of viral entry into cells is a conceptually sound antiviral strategy. The "dance" between SARS-CoV and its host cell involves binding, conformational change of S2 and membrane fusion, all of which are targets for therapy.

Chimeric Protein that Neutralizes SARS-CoV Spike Protein

^{*}Address correspondence to this author at the Department of Pathology, Princess Margaret Hospital, Hong Kong; Tel: 1,661-889-8218; Fax: 1,661-885-5297; E-mail: tommy.tong@yahoo.com

The first step of viral entry is attachment of viral surface molecule with its cellular receptor. Soluble decoy receptors that saturate these viral molecules could be used to prevent viral binding to cells.



Fig. (1). SARS-CoV genome. A 29-nucleotide stretch is deleted in the humanized strain. Compare with that of bat and civet strains at the bottom.

	Mechanism	IC_{50}^{1}/EC_{50}^{2}	CC ₅₀ ³	SI ⁴	Reference
HR121	Heptad repeat (Entry inhibitor)	4.13 µM	-	-	29
HR212	Heptad repeat (Entry inhibitor)	0.95 µM	-	-	29
MDL28170	Cathepsin L inhibitor (Entry inhibitor)	2.5 nM	-	-	40
Chloroquine	Entry inhibitor	8.8 +/- 1.2 µM	261.3 +/- 14.5 μM	30	44
Chloroquine	Entry inhibitor	4.4 +/- 1.0 μM	-	-	45
FP-21399	? Entry inhibitor	Low µM	-	-	53
AG7088	Protease inhibitor	Not effective at 10 µM	-	-	53
KZ7088	Protease inhibitor	-	-	-	54, 55
Ritonavir	Protease inhibitor	Not effective at 50 µM	-	-	53
Saquinavir	Protease inhibitor	Not effective at 50 µM	-	-	53
Lopinavir	Protease inhibitor	50 µM	-	-	53
Lopinavir-like compounds 26-36	Protease inhibitor	23-40 µM	-	-	53

Table 1. Properties of Some SAKS-Cov Anti-Infectiv	able 1.	. Properties of	Some SARS-CoV	Anti-Infectives
--	---------	-----------------	---------------	-----------------

(Table 1) Contd....

	Mechanism	IC_{50}^{1}/EC_{50}^{2}	CC ₅₀ ³	SI^4	Reference
Lopinavir	Protease inhibitor	l μg/ml (with ribavirn 6.25 μg/ml)	-	-	67
MAC-5576	Protease inhibitor	05. +/- 0.3 μM	-	-	57
MAC-8120	Protease inhibitor	4.3 +/- 0.5 μM	-	-	57
MAC-13985	Protease inhibitor	7 +/- 2 µM	-	-	57
MAC-22272	Protease inhibitor	2.6 +/- 0.4 µM	-	-	57
MAC-30731	Protease inhibitor	7 +/- 3 µM	-	-	57
Tannic acid	Protease inhibitor	3 µM	-	-	61
TF2B	Protease inhibitor	7 μM	-	-	61
TF3	Protease inhibitor	<10 µM	-	-	61
Hesperetin	Protease inhibitor	8.3 µM	-	-	62
Ribavirin	Polymerase inhibitor	0.5-5 mg/ml	0.2-1 mg/ml	<1	74
-D-N4-hydroxycytidine	Polymerase inhibitor	5 µM	50 µM	10	78
Aurintricarboxylic acid	Polymerase inhibitor	0.2 mg/ml	37.5 mg/ml	187	83
Valinomycin	?	0.85 µM	68 µM	80	53
Reserpine	?	3.4 µM	25 µM	7.3	53
Reserpine derivatives (compounds 19-24)	?	<100 µM	-	-	53
Aescin	?	6.0 µM	15 µM	2.5	53
Bananins	Helicase inhibitor	<10 µM	>300 µM	>30	88
Glycyrrhizin derivatives (compounds 6, 6, 17 & 18)	?	<100 µM	-	-	53
Glycyrrhizin	?	300 mg/l	>20,000 mg/l	>67	117
Niclosamide	Unknown	1-3 µM	250 µM	-	116
Calpain inhibitor VI	?	3 µM (EC ₉₀) virus yield reduction assay	-	-	78
Calpain inhibitor III	?	15 μM (EC ₉₀) virus yield reduction assay	-	-	78

1 - IC_{50} – Concentration of a drug that is required for 50% inhibition of viral replication *in vitro*.

2 - EC_{50} – Plasma concentration required for obtaining 50% of the maximal effect *in vivo*.

3 - $CC_{\rm 50}$ - Cytotoxic concentration that reduced cell viability to 50%.

4 - SI (Selectivity index) = CC_{50}/EC_{50} .

In HIV infection, more than a decade of research has shown that unmodified decoy receptors are not sufficiently potent [16-18]. The strategy quickly "evolved" into a novel HIV entry inhibitor composed of chimeric proteins, such as CD4-IgG [19-21]. This chimeric molecule has multiple binding regions for HIV-1 gp41 and is currently being tested in human subjects, e.g. ClinicalTrials.gov, identifier NCT 00000876. The chimera overcomes the problems of soluble CD4, such as low neutralizing activity, enhancement of viral infection, and short half-life *in vivo*. Jacobson *et al.* evaluated PRO 542 [22], a CD4-IgG2, in HIV-infected adults in a phase 1 study and reported reductions in plasma HIV RNA and plasma viremia with no dose-limiting toxicities [23]. In another phase 1/2 clinical trial in children with HIV-I infection, PRO 542 was again shown to be well-tolerated besides reducing the viral burden [24].

In a similar fashion, engineered multivalent soluble ACE2 (sACE2)-immunoglobulin might also be efficacious

in neutralizing SARS-CoV [25]. sACE2 can conceivably be improved by using residues 90-93 of civet ACE2 [26].

Membrane Fusion Inhibitors

SARS-CoV shares a similar mechanism with HIV-1 in achieving membrane fusion between virus and host cell. Thus, heptad repeats (HR1 and HR2) located in the S2 domain of SARS-CoV spike protein, oligomerizes to form a six-helix bundle after attachment to ACE2. Spike protein heptad repeat-derived peptides have therefore been predicted [27] and recently shown to inhibit SARS-CoV infection of Vero cells [28]. Further efforts resulted in stable recombinant proteins containing HR1 and HR2, having potent inhibitory activities (HR121 and HR212; IC₅₀ values of 4.13 and 0.95 μ M, respectively) on entry of the HIV/SARS pseudoviruses [29]. These proteins are also more economical to produce than synthetic peptides. However, they will need to be administered parenterally.

Cathepsin L Inhibitors

Cathepsins are host intracellular enzymes belonging to the papain family of cysteine proteases, of which there are over a dozen types. Most of them are activated by the low pH environments of lysosomes and endosomes, in which they function. Better known cathepsins include type A, required to stabilize sialidase and -galactosidase; type B, involved in activation of tissue plasminogen and cancer metastasis [30, 31]; type D, involved in mediating apoptosis [32]; type H, expressed in renal oncocytomas but not carcinomas [33]; type K, involved in degradation of type I collagen (mutated in congenital bone disorder) and cancerinduced osteolysis [34]; type L, required for degradation of li in cortical thymic epithelial cells but not marrow-derived antigen-presenting cells [35] and probably participating in malignant transformation [36]; type N, also with collagenolytic activity [37]; and type S, which is inducible by interferon (IFN)- in MHC-class II expressing cells and is pivotal in the maturation and peptide-binding competency of class II molecules [38].

To infect cells after SARS-CoV binds to its receptor ACE2, cathepsin L was found to be necessary for an as yet incompletely understood endosomal step [39]. This cellular protease is not however, required by human coronavirus NL63, which also utilizes ACE2 as receptor.

Specific inhibitors of cathepsin L has now been shown to prevent SARS-CoV infection *in vitro* [39, 40]. For example, MDL28170 (calpain inhibitor III) had an IC₅₀ of 2.5 nM on substrate cleavage and efficient inhibition of SARS-CoV replication *in vitro*.

Cathepsin L inhibitor, 4-amino-azepan-3-one compounds (Fig. 2) described in US patent application 20040192674 [41] claims activity in rheumatoid arthritis and prevention of cancer metastasis, among other indications. It would be worthwhile to determine if it possesses anti-viral activity against SARS-CoV.

US20030229226 [42] also teaches small molecule compounds with inhibitory activity against cathepsins K and L.

In addition to small molecules, cellular proteins such as cathepsin L are also susceptible to RNAi-based intervention.



Fig. (2). A representative 4-amino-azepan-3-one compound. Formula A (quinoline-6-carboxylic acid {(*S*)-naphthylen-2-yl-1-[(*S*)-oxo-1-(pyridine-2-sulfonyl)-azepan-4-yl carbamoyl]-ethyl}amide). It inhibits cathepsin L and may prevent entry of SARS-CoV.

Chloroquine

A potentially useful old drug is Chloroquine (Fig. 3), being tested for its anti-HIV effect in clinical trials [43]. It is a 9-aminoquinoline discovered by German chemist Hans Andersag in 1934 and used for the treatment of malaria, amebiasis, and autoimmune diseases such as rheumatoid arthritis. It has a high selectivity index of 30 against SARS-CoV in in vitro studies [44]. At 10 µM concentration, achievable in vivo at dosages used for malaria prophylaxis and treatment, viral inhibition was total by immunofluorescence assay [45]. Chloroquine increases endosomal pH, which explains its similar efficacy as ammonium chloride, another lysosomotropic agent, when given up to 5 hours after infection of cell culture by SARS-CoV [44,45]. However, it is also effective when given before viral inoculation onto cell culture, probably due to its interference with terminal glycosylation of ACE2 [44].

Recently, novel synthetic organometallic compounds closely mimicking hydroxychloroquine were found to have selective effect on SARS-CoV. The cytotoxic effects as expected, are less than the parent ferroquine compound [46].



Fig. (3). Chloroquine (7-Chloro-4-(4-Diethylamino-1-Methylbutylamino) Quinoline). This anti-malarial probably has multiple actions that prevent SARS-CoV from entering cells.

II. SARS-COV PROTEASES AS PROMISING DRUG TARGET

SARS-CoV 3CLpro is a promising drug target because it is essential to the formation of a functional replication complex [47-49]. With the availability of the SARS-CoV genome [10,11,50], a homology model of SARS-CoV chymotrypsin-like protease (main protease, also called 3CLpro) was constructed, providing a basis for the design of anti-SARS drugs. This model is based on the crystal structures of HCoV-229E main protease (M^{PRO}) and TGEV M^{PRO} in complex with AG7088 [51]. Functional conservation among the coronaviruses suggests that a drug with Gln

(Ser,Ala,Gly) specificity (denotes cleavage site) against SARS 3CLpro may also have activity against the other members [52]. This becomes highly relevant with the recent discovery of several SARS-CoV-like viruses in the wild [6, 7]. This functional conservation was demonstrated by structural homology studies [51].

These results led to the speculation that an anti-rhinoviral drug already in clinical trial, AG7088 (Fig. 4), might be useful against SARS-CoV. It was subsequently found not to have *in vitro* activity against the virus at a concentration of 10 μ M [53]. However, its derivative KZ7088 (Fig. 5) interacts specifically with the active site of SARS-CoV 3CLpro through six hydrogen bonds [54]. Based on the atomic coordinates obtained by docking KZ7088 with the enzyme's active site, a pharmacophore virtual screening narrowed down the list of compounds worthy of further experiments to 0.03% of the 3.6 million screened [55].



Fig. (4). AG7088. This molecule is being tested in clinical trials against rhinovirus. However, it has no *in vitro* activity against SARS-CoV at a concentration of 10μ M.



Fig. (5). KZ7088. A derivative of AG7088, KZ7088 interacts specifically with the active site of SARS-CoV 3CLpro.

Recently, proteomic technologies were employed to assist in peptide-based screening of 3CLpro substrate specificity [56]. This information will further assist in drug design. Moreover, this approach using matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS) is readily adaptable to study the substrate specificity of other proteases in a high throughput manner.

Compounds with Activity Against SARS Main Protease (3CLpro)

Using a quenched fluorescence resonance energy transfer assay (Abz-peptide-Nitrotyrosine) and screening 50,000 drug-like small molecules, Canadian scientists discovered five new molecules with 3CLpro IC₅₀ in the range of 0.5-7 μ M (MAC-5576, -8120, -13985, -22272, -30731) [57].

Recently, benzotriazole esters, intermediates in the synthesis of lopinavir, were found to have k_{inact} (maximal rate of enzyme inactivation) of 0.0011 sec⁻¹ and a K_{I} (inhibitor concentration that supports half the maximal rate of inactivation) of 7.5 nM against 3CLpro [58].

In addition, a group of dicyclic or multi-cyclic compounds with inhibitory activity against SARS-CoV main protease were disclosed in US patent publication 200600-19967 [59]. An example is compound (18) (Fig. 6).



Fig. (6). Compound (18) with inhibitory activity against SARS-CoV 3CLpro (US20060019967).

Natural Substances with Activity Against SARS Main Protease (3CLpro)

Theaflavins from black tea has activity against bovine rotavirus and coronavirus as shown by *in vitro* studies. The EC_{50} against coronavirus was 34.7 micrograms/ml [60].

Substances in black tea with activity against SARS-CoV were discovered as part of a large-scale screening process involving 720 natural products. These include simple and complex oxygen heterocycles, alkaloids, sequiterpenes, diterpenes, pentacyclic triterpenes, and sterols [61]. Assaying for inhibitory activity against 3CLpro proteolytic activity using HPLC, tannic acid and TF2B were identified as having IC₅₀ at concentrations <10 μ M. Additional experiments using well-known ingredients in tea resulted in the findings that the gallate group-containing TF2B and TF3 have more potent 3CLpro-inhibitory activity than TF1 (Fig. 7). Thus, tannic acid, TF2B and TF3 join the list of compounds that require evaluation for activity in cell culture.

In another study, *Isatis indigotica* root extract, major compounds from *I. indigotica* root, and several plant-derived phenolic compounds were tested for anti-SARS-CoV 3CLpro inhibitory activity in *in vitro* assays and cell cultures [62]. Cleavage assays with 3CLpro demonstrated that IC_{50} values were in μ M ranges for *I. indigotica* root extract, indigo, sinigrin, aloe emodin and hesperetin. Hesperetin (Fig. **8**) dose-dependently inhibited cleavage activity of the 3CLpro, in which the IC_{50} was 8.3 μ M in cell-based assay. Thus hesperetin needs to be further investigated.



Fig. (7). Chemical structures of TF1, TF2B and TF3.

Papain-Like Cysteine Protease Inhibitors

SARS-CoV Papain-like cysteine protease is a Zn-ribboncontaining proteinase and conserved with the corresponding PL2pro of other coronaviruses. SARS-CoV does not have a



Fig. (8). Hesperetin is a bioflavonoid. It is the aglycone of hesperidin, found in citrus fruits. Hesperetin dose-dependently inhibited cleavage activity of 3CLpro in cell-based assay.

homolog of PL1pro that is present in other family members. PL2pro probably took over the function of cleaving the N-terminal portions of pp1a and pp1ab [12,63]. Comparative studies of coronaviral PLpro conservation suggested a link with substrate specificities, which SARS-CoV PL2pro demonstrates. This narrow specificity for substrates is an Achilles' heel that may be exploited for drug development [63].

Clinical Experience with Protease Inhibitors

During the epidemic clinicians in Guangzhou observed that HIV-positive patients on HAART appear to be protected against SARS [64,65]. In Hong Kong, the utility of lopinavir-ritonavir was investigated in a multi-center retrospective matched cohort study as initial and rescue therapy for SARS [66]. Patients who received this therapy as initial treatment for SARS had better outcome (reduced death and intubation rate) compared with an uncontrolled group, with lower rate of use of methylprednisolone at a lower mean dose. The results were similar to another report of a subset of those patients treated by the same senior researcher [67]. These clinical trials are in agreement with structural studies that predicted the utility of lopinavir, ritonavir, niclosamide and promazine against 3CL pro [68], with lopinavir and nelfinavir also showing in vitro activity [67,69,70].

III. SARS-COV POLYMERASE AS DRUG TARGET

Widespread interest in coronaviruses is relatively recent. As a result, there is little experimental data on the characteristics of coronaviral RNA-dependent RNA polymerase (RdRp) and a consequent lack of inhibitors for this enzyme. The situation is very different for hepatitis B and C, HIV and herpes viruses, where polymerase inhibitors are very successful clinically. SARS-CoV RdRp, which is very important in the viral life cycle, is therefore high on the list of drug targets [71].

Ribavirin

Ribavirin (1-(-D-Ribofuranosyl)-1H-1,2,4-triazole-3carboxamide) was used extensively during the epidemic (Fig. 9). Derived from D-ribose, it is a long half-life purine nucleoside analog that interacts with viral RNA polymerases, as well as having other activities, such as inhibition of cellular inosinate (IMP) dehydrogenase [72]. In treatment of HCV, ribavirin is thought to act by inhibition of IMP dehydrogenase and by enhancement of Th1 activities [73].

For SARS-CoV however, at a low selectivity index (SI; SI=CC₅₀/EC₅₀) of <1 [74], coupled with an absence of demonstrable clinical benefit in uncontrolled series [75,76], the role of ribavirin in treatment is in doubt. The side effects include teratogenicity and a dose-dependent but reversible hemolytic anemia [77]. The N-terminal domain of SARS-CoV nsp14 is homologous to 3'-to-5' exonulcease (ExoN) and may perform RNA proofreading, repair and/or recombination [12]. This unusual capability among RNA viruses may be responsible for the failure of ribavirin in SARS-CoV therapy. Recently, data emerged that ribavirin and other IMP dehydrogenase inhibitors enhance lung infection in a BALB/c mice model [78]. The continued use of ribavirin alone is not recommended [76,77], unless perhaps in combination with an ExoN-inhibitor and supported by experimental results.



Fig. (9). Ribavirin. Ribavirin has low selectivity index (<1) against SARS-CoV.

Other Nucleotides, Nucleosides, and Nucleoside Analogs

Despite the attractiveness of this group of drugs, -D-N4hydroxycytidine (Fig. **10**) is the only nucleoside analog among 26 tested that is selective and has an ED₉₀ of 6 μ M by virus yield-reduction assay [79]. Its CC₅₀ and EC₅₀ were 50 and 5 μ M, respectively (SI=10). It was earlier found to have selective activity against hepatitis C virus [80-82].



Fig. (10). -D-N4-hydroxycytidine. This is the only member among 26 nucleoside analogs tested that has selective activity against SARS-CoV.

Other Drugs that Inhibit SARS-CoV Polymerase

Aurintricarboxylic acid (ATA) is a general inhibitor of nucleases found recently to be more potent than IFNagainst SARS-CoV [83]. Molecular docking studies suggest that it inhibits SARS-CoV RdRp by binding to a region in the palm domain (754-766), where two of the three catalytic residues (Asp 760, Asp 761) are located [84].

IV. SARS-COV HELICASE AS DRUG TARGET

This enzyme is another viral enzyme that is worth investigating for drug development. Earlier work has revealed that the viral enzyme unwinds DNA as well as RNA. This property facilitates the development of high-throughput DNA-based helicase assays, which will facilitate the search for inhibitors [63]. That the effort may be worthwhile can be seen in the success of helicase inhibitors currently being developed for herpes viruses [85,86] and HCV [87].

Recently, several bananins (pyridoxal-conjugated trioxaadamantanes), including iodobananin, bananin (Fig. 11), eubananin (Fig. 12) and vanillinbananin (Fig. 13) were found to non-competitively inhibit the ATPase activity of SARS-CoV helicase with IC₅₀ values in the range of 0.5-3 μ M [88]. In cell culture, bananin has an EC₅₀ of <10 μ M and a CC₅₀ of >300 μ M (SI of >30). Steric hindrance around the pyridoxal ring of some bananins (ansabananin [Fig. 14] and adeninobananin) appears to explain why they have no anti-SARS-CoV activity. Surprisingly, bananins may aggravate infection when given prophylactically.



Fig. (11). Bananin.



Fig. (12). Eubananin.



Fig. (13). Vanillinbananin.



Fig. (14). Ansabananin.

V. INTERFERONS AND INTERFERON INDUCERS

Interferons in SARS

Like most metazoan viruses, SARS-CoV targets the interferon (IFN) signaling pathway [89-91], highlighting the evolutionary importance of IFN against viral infections. Not unexpectedly, evidences of IFN efficacy *in vitro* were readily established during the epidemic.

The type-I IFNs (/) but not type-II INF () were found to inhibit SARS-CoV infection and replication [92]. Natural IFN- and IFN- have more potent *in vitro* activity than recombinant IFN- [93]. Alferon N Injection is the only approved natural, multi-species, alpha-interferon available in the US. *in vitro* Studies demonstrated specific anti-SARS-CoV activity in Vero 76 cell culture. Alferon inhibited SARS-CoV at an EC₅₀ of 5,696 +/- 1,703 (SEM) IU/ml (visual) and 10,740 +/- 5,161 (SEM) IU/ml (neutral red). Viral load reduction by one log₁₀ was 78,000 +/- 22,000 (SEM) IU/ml [94].

The *in vivo* activity of interferons were confirmed in macaques, which were protected from SARS-CoV by prophylactic pegylated IFN- [95]. Postexposure treatment also produced measurable antiviral efficacy, supporting the rationale of employing IFN for prophylaxis or treatment of SARS. In humans, uncontrolled clinical trials have been reported [74,96-100]. No double-blind randomized controlled trial has yet been conducted.

Interferon Inducers

Substances that induce IFN production by dendritic cells and peripheral blood mononuclear cells, such as CpG oligodeoxynucleotide (BW001) yielded supernatant that protected Vero cells from SARS-CoV infection [101].

Hemispherx biopharma recently filed a patent application [94], for treatment of acute and severe viral infections that includes natural human alpha interferons and Ampligen. *in vitro* Cytopathic effect-prevention data on influenzaviruses using various combinations of interferon, ribavirin, oseltamivir and Ampligen were presented.

Ampligen ($rI_n.r(C_{12}U)_n$, Poly APoly U or $rI_nr(C_{29},G)_n$, in which r is ribo) is a mismatched derivative of double-stranded RNAs. It was recently reviewed by De Clercq with

the interferons [102]. Mice given Ampligen alone were protected from coxsackie B3 virus-induced myocarditis [103] and flavirus-induced encephalitis [104] but offered only limited protection against lethal pichinde virus challenge [105]. When given with interferon, Ampligen amplifies its effects. It also has synergistic activities with most anti-retrovirals [106] and has generated data in clinical trials [107]. No data is available for SARS-CoV.

VI. MONOCLONAL ANTIBODIES AS ANTIVIRALS

Neutralizing convalescent or engineered antibodies have therapeutic potential in SARS-CoV infection [108]. Neonatal respiratory syncytial virus infections have been prevented by prophylactic administration of MEDI-493, a humanized monoclonal antibody [109].

The SARS animal model ferret, was protected by prior administration of monoclonal antibodies [110]. Moreover, convalescent serum has been used in SARS patients and mice without ill effect, with mice showing measurable antiviral activity against SARS-CoV [111-113].

One group developed an improved B-cell immortalization technique using CpG 2006 (a CpG oligonucleotide) as polyclonal B-cell activator. Employing irradiated allogeneic mononuclear cells, Epstein-Barr virus, and CpG 2006, they interrogated the B-cell memory repertoire of an immune SARS patient. S3.1, a neutralizing antibody from one stable B-cell clone was found to protect mice lungs from SARS-CoV challenge [112]. A total of thirty five monoclonal neutralizing antibodies were isolated in this study. The drawback of the method is that convalescent patients are required.

Another approach uses non-immune human antibody libraries. Eight recombinant human single-chain variable region fragments (scFvs) against the RBD of S protein were identified from a vast library. 80R IgG1, a monoclonal antibody engineered from one such fragment possesses potent neutralizing activity in *in vitro* and animal studies [114,115].

VII. OTHER SARS-COV ANTI-INFECTIVES

Mannose-binding lectin, a component of the innate immune system, proves to play a role in preventing SARS-CoV infection. Deficiency, usually constitutional, is associated with SARS. Therapeutic and prophylactic replenishment needs to be further investigated [116]. Calpain inhibitors have shown some promise, although one of them (calpain inhibitor III) appears to work primarily on cathepsin L rather than the purported main protease 3CLpro [40,79].

Niclosamide

Discovering novel drugs by chemical screening is one strategy but others adopted a strategy of screening old drugs for novel antiviral activities against SARS-CoV [68].

Such efforts resulted in the identification of niclosamide (2',5-dichloro-4'-nitrosalicylanilide) (Fig. 15), an antihelminthic agent, as having potent activity [68,117]. Vero E6 cells preincubated with drug at 10 μ M concentration for 1 hour (also effective 3 hours after infection) and infected by SARS-CoV at an MOI (multiplicity of infection) of 0.1 were



Fig. (15). Niclosamide. This antihelminthic agent has potent *in vitro* activity against SARS-CoV.

observed for protection against CPE (cytopathic effect). Immunofluorescent assay determined the EC_{50} to be between 1 and 3 μ M, whereas the CC_{50} (cytotoxic concentration that reduced cell viability to 50%) was 250 μ M (at 48 hours incubation). The mechanism of viral inhibition by Niclosamide is not dependent upon inhibition of entry or anti-3 CL^{PRO} activity and remains to be determined.

Glycyrrhizin

Another such compound is glycyrrhizin [118] and its derivatives [53,119]. Glycyrrhizin (Fig. 16) a triterpenoid saponin found in *Glycyrrhiza glabra* (licorice). It stands out among the inosine monophosphate decarboxylase inhibitors (ribavirin and mycophenolic acid) and orotidine monophosphate decarboxylase inhibitors (6-azauridine and pyrazofurin) as having a high selectivity index (SI=CC₅₀/EC₅₀) of 67 against SARS-CoV. Its mechanisms of action are uncertain and may be due to its effects on protein kinase C (cellular signaling pathway), AP-1, NF- B (transcription factors), and upregulation of inducible nitrous oxide synthase and increased production of nitrous oxide by macrophages. Experimental support of the latter mode of action was provided by the induction of nitrous oxide synthase activity by glycyrrhizin and the fact that addition of nitrous oxide donor (DETA NONOate) inhibits viral replication in Vero E6 cells [118]. Moreover, its effect on lowering plasma membrane fluidity and hence impeding viral entry, is consistent with its observed broad antiviral activity [120]. Side effects include hypertension and hypokalemia in some patients after prolonged treatment.



Fig. (16). Glycyrrhizin. Several studies demonstrated a high selectivity index of this agent against SARS-CoV. Its anti-SARS-CoV actions are likely to be multiple.

Valinomycin

Valinomycin (Fig. **17**), a dodecadepsipeptide (a macrocyclic molecule made of twelve alternating amino acids) potassium transporter obtained from the cells of several *Streptomyces* strains, one of them *S. Tsusimaensis*, has EC₅₀, based on ELISA, of 0.85 μ M (SI = 80). The mode of antiviral action is unclear [53]. In the same study, FP-21399, and some saponins have also being identified as being highly effective and worthy of further study.



Fig. (17). Valinomycin.

CURRENT & FUTURE DEVELOPMENTS

It is apparent from this survey that anti-SARS-CoV drug development is advancing on many fronts and that many molecules have been screened. It is also apparent that much needs to be elucidated, in particular the *in vivo* activity of these agents in animal studies. However, some of these agents, particularly traditional remedies have well-known safety profiles and should receive priority for further developments [69]. As antivirals come into use, the issue of drug resistance will emerge, as in the case of neuraminidase inhibitor resistance in influenzavirus. Viral escape from drug activity is a virtual certainty and needs to be anticipated and monitored.

Also needing further development are RNA interferencebased [121-127] and anti-sense therapies [128], as exemplified by US20060063150 [129], which employs uncharged highly stable morpholino oligonucleotides. The specific delivery of such cargoes into target cells has been a challenge. Recently, Song *et al.* gave us hope that an antibody-mediated delivery via cell-surface receptors may revolutionize this field [130,131].

The rapid accumulation of anti-SARS-CoV medications is a welcome sign of strong basic research. We are now in bad need for paradigm shifts that would lead to more vigorous, but unglamorous, and non-Nobel-winning translational research as well as to inculcate a business ethic of investing in money-losing products that nevertheless might still benefit the pharmaceutical industry in unimaginable ways.

REFERENCES

- Peiris JS, Lai ST, Poon LL, *et al.* Coronavirus as a possible cause of severe acute respiratory syndrome. Lancet 2003; 361: 1319-25.
- Peiris JS, Guan Y, Yuen KY. Severe acute respiratory syndrome. Nat Med 2004; 10: S88-97.
- [3] Peiris JS, Yuen KY, Osterhaus AD, Stohr K. The severe acute respiratory syndrome. N Engl J Med 2003; 349: 2431-41.
- [4] Drosten C, Preiser W, Gunther S, Schmitz H, Doerr HW. Severe acute respiratory syndrome: identification of the etiological agent. Trends Mol Med 2003; 9: 325-27.
- [5] Ksiazek TG, Erdman D, Goldsmith CS, et al. A novel coronavirus associated with severe acute respiratory syndrome. N Engl J Med 2003; 348: 1953-66.
- [6] Lau SK, Woo PC, Li KS, et al. Severe acute respiratory syndrome coronavirus-like virus in Chinese horseshoe bats. Proc Natl Acad Sci USA 2005; 102: 14040-45.
- [7] Li W, Shi Z, Yu M, et al. Bats Are Natural Reservoirs of SARS-Like Coronaviruses. Science 2005; 310: 676-79.
- [8] Heymann DL. The international response to the outbreak of SARS in 2003. Philos Trans R Soc Lond B Biol Sci 2004; 359: 1127-29.
- [9] Chinese SMEC. Molecular evolution of the SARS coronavirus during the course of the SARS epidemic in China. Science 2004; 303: 1666-69.
- [10] Marra MA, Jones SJ, Astell CR, et al. The Genome sequence of the SARS-associated coronavirus. Science 2003; 300: 1399-404.
- [11] Rota PA, Oberste MS, Monroe SS, et al. Characterization of a novel coronavirus associated with severe acute respiratory syndrome. Science 2003; 300: 1394-99.
- [12] Snijder EJ, Bredenbeek PJ, Dobbe JC, et al. Unique and conserved features of genome and proteome of SARScoronavirus, an early split-off from the coronavirus group 2 lineage. J Mol Biol 2003; 331: 991-1004.
- [13] Li W, Wong SK, Li F, *et al.* Animal origins of the severe acute respiratory syndrome coronavirus: Insight from ACE2-S-protein interactions. J Virol 2006; 80: 4211-19.
- [14] De Clercq E. Molecular targets for antiviral agents. J Pharmacol Exp Ther 2001; 297: 1-10.
- [15] Holmes KV. SARS coronavirus: a new challenge for prevention and therapy. J Clin Invest 2003; 111: 1605-09.
- [16] Traunecker A, Luke W, Karjalainen K. Soluble CD4 molecules neutralize human immunodeficiency virus type 1. Nature 1988; 331: 84-86.
- [17] Deen KC, McDougal JS, Inacker R, et al. A soluble form of CD4 (T4) protein inhibits AIDS virus infection. Nature 1988; 331: 82-84.
- [18] Fisher RA, Bertonis JM, Meier W, et al. HIV infection is blocked in vitro by recombinant soluble CD4. Nature 1988; 331: 76-78.
- [19] Byrn RA, Sekigawa I, Chamow SM, et al. Characterization of in vitro inhibition of human immunodeficiency virus by purified recombinant CD4. J Virol 1989; 63: 4370-75.
- [20] Capon DJ, Chamow SM, Mordenti J, et al. Designing CD4 immunoadhesins for AIDS therapy. Nature 1989; 337: 525-31.
- [21] Traunecker A, Schneider J, Kiefer H, Karjalainen K. Highly efficient neutralization of HIV with recombinant CD4immunoglobulin molecules. Nature 1989; 339: 68-70.
- *[22] Maddon, P.J., Beaudry, G.A.: US20016187748 (2001).
- [23] Jacobson JM, Lowy I, Fletcher CV, et al. Single-dose safety, pharmacology, and antiviral activity of the human immunodeficiency virus (HIV) type 1 entry inhibitor PRO 542 in HIVinfected adults. J Infect Dis 2000; 182: 326-29.
- [24] Shearer WT, Israel RJ, Starr S, et al. Recombinant CD4-IgG2 in human immunodeficiency virus type 1-infected children: phase 1/2 study. The Pediatric AIDS Clinical Trials Group Protocol 351 Study Team. J Infect Dis 2000; 182: 1774-79.
- [25] Dimitrov DS. The secret life of ACE2 as a receptor for the SARS virus. Cell 2003; 115: 652-53.
- [26] Li W, Zhang C, Sui J, et al. Receptor and viral determinants of SARS-coronavirus adaptation to human ACE2. EMBO J 2005; 24: 1634-43.
- [27] Kliger Y, Levanon EY. Cloaked similarity between HIV-1 and SARS-CoV suggests an anti-SARS strategy. BMC Microbiol 2003; 3: 20.

- [28] Bosch BJ, Martina BE, Van Der Zee R, et al. Severe acute respiratory syndrome coronavirus (SARS-CoV) infection inhibition using spike protein heptad repeat-derived peptides. Proc Natl Acad Sci USA 2004; 101: 8455-60.
- [29] Ni L, Zhu J, Zhang J, Yan M, Gao GF, Tien P. Design of recombinant protein-based SARS-CoV entry inhibitors targeting the heptad-repeat regions of the spike protein S2 domain. Biochem Biophys Res Commun 2005; 330: 39-45.
- [30] Kobayashi H, Schmitt M, Goretzki L, et al. Cathepsin B efficiently activates the soluble and the tumor cell receptorbound form of the proenzyme urokinase-type plasminogen activator (Pro-uPA). J Biol Chem 1991; 266: 5147-52.
- [31] Szpaderska AM, Frankfater A. An intracellular form of cathepsin B contributes to invasiveness in cancer. Cancer Res 2001; 61: 3493-500.
- [32] Deiss LP, Galinka H, Berissi H, Cohen O, Kimchi A. Cathepsin D protease mediates programmed cell death induced by interferon-gamma, Fas/APO-1 and TNF-alpha. EMBO J 1996; 15: 3861-70.
- [33] Castren JP, Kamel DE, Nurmi MJ, Collan YU. Cathepsin H expression distinguishes oncocytomas from renal cell carcinomas. Anticancer Res 2000; 20: 537-40.
- [34] Gelb BD, Shi GP, Chapman HA, Desnick RJ. Pycnodysostosis, a lysosomal disease caused by cathepsin K deficiency. Science 1996; 273: 1236-38.
- [35] Nakagawa T, Roth W, Wong P, et al. Cathepsin L: critical role in Ii degradation and CD4 T cell selection in the thymus. Science 1998; 280: 450-53.
- [36] Kane SE, Gottesman MM. The role of cathepsin L in malignant transformation. Semin Cancer Biol 1990; 1: 127-36.
- [37] Maciewicz RA, Etherington DJ. A comparison of four cathepsins (B, L, N and S) with collagenolytic activity from rabbit spleen. Biochem J 1988; 256: 433-40.
- [38] Riese RJ, Wolf PR, Bromme D, et al. Essential role for cathepsin S in MHC class II-associated invariant chain processing and peptide loading. Immunity 1996; 4: 357-66.
- [39] Huang IC, Bosch BJ, Li F, et al. SARS coronavirus, but not human coronavirus NL63, utilizes cathepsin L to infect ACE2expressing cells. J Biol Chem 2006; 281: 3198-203.
- [40] Simmons G, Gosalia DN, Rennekamp AJ, Reeves JD, Diamond SL, Bates P. Inhibitors of cathepsin L prevent severe acute respiratory syndrome coronavirus entry. Proc Natl Acad Sci USA 2005; 102: 11876-81.
- *[41] Marquis, R.W.: US20040192674 (2004).
- *[42] Okamoto, O., Falgueyret, J., Oballa, R., Prasit, P., Rydzewski, R.: US20030226226 (**2003**).
- [43] Savarino A, Boelaert JR, Cassone A, Majori G, Cauda R. Effects of chloroquine on viral infections: an old drug against today's diseases? Lancet Infect Dis 2003; 3: 722-27.
- [44] Keyaerts E, Vijgen L, Maes P, Neyts J, Van Ranst M. *in vitro* Inhibition of severe acute respiratory syndrome coronavirus by chloroquine. Biochem Biophys Res Commun 2004; 323: 264-68.
- [45] Vincent MJ, Bergeron E, Benjannet S, *et al.* Chloroquine is a potent inhibitor of SARS coronavirus infection and spread. Virol J 2005; 2: 69.
- [46] Biot C, Daher W, Chavain N, et al. Design and synthesis of hydroxyferroquine derivatives with antimalarial and antiviral activities. J Med Chem 2006; 49: 2845-49.
- [47] Bacha U, Barrila J, Velazquez-Campoy A, Leavitt SA, Freire E. Identification of novel inhibitors of the SARS coronavirus main protease 3CLpro. Biochemistry 2004; 43: 4906-12.
- [48] Lee TW, Cherney MM, Huitema C, *et al*. Crystal structures of the main peptidase from the SARS coronavirus inhibited by a substrate-like Aza-peptide epoxide. J Mol Biol 2005; 353: 1137-51.
- [49] Martina E, Stiefl N, Degel B, et al. Screening of electrophilic compounds yields an aziridinyl peptide as new active-site directed SARS-CoV main protease inhibitor. Bioorg Med Chem Lett 2005; 15: 5365-69.
- [50] Leung FC. Hong Kong SARS sequence. Science 2003; 301: 309-10.
- [51] Anand K, Ziebuhr J, Wadhwani P, Mesters JR, Hilgenfeld R. Coronavirus main proteinase (3CLpro) structure: basis for design of anti-SARS drugs. Science 2003; 300: 1763-67.

- [52] Yang H, Xie W, Xue X, et al. Design of wide-spectrum inhibitors targeting coronavirus main proteases. PLoS Biol 2005; 3: e324.
- [53] Wu CY, Jan JT, Ma SH, et al. Small molecules targeting severe acute respiratory syndrome human coronavirus. Proc Natl Acad Sci USA 2004; 101: 10012-17.
- [54] Chou KC, Wei DQ, Zhong WZ. Binding mechanism of coronavirus main proteinase with ligands and its implication to drug design against SARS. Biochem Biophys Res Commun 2003; 308: 148-51.
- [55] Sirois S, Wei DQ, Du Q, Chou KC. Virtual screening for SARS-CoV protease based on KZ7088 pharmacophore points. J Chem Inf Comput Sci 2004; 44: 1111-22.
- [56] Chu LH, Choy WY, Tsai SN, Rao Z, Ngai SM. Rapid peptidebased screening on the substrate specificity of severe acute respiratory syndrome (SARS) coronavirus 3C-like protease by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry. Protein Sci 2006; 15: 699-709.
- [57] Blanchard JE, Elowe NH, Huitema C, et al. High-throughput screening identifies inhibitors of the SARS coronavirus main proteinase. Chem Biol 2004; 11: 1445-53.
- [58] Wu CY, King KY, Kuo CJ, et al. Stable benzotriazole esters as mechanism-based inactivators of the severe acute respiratory syndrome 3CL protease. Chem Biol 2006; 13: 261-68.
- *[59] Wu, S.Y., Hsieh, H.P., Hsu, T.A., Lu, I.L.: US20060019967 (2006).
- [60] Clark KJ, Grant PG, Sarr AB, et al. An in vitro study of the aflavins extracted from black tea to neutralize bovine rotavirus and bovine coronavirus infections. Vet Microbiol 1998; 63: 147-57.
- [61] Chen CN, Lin CP, Huang KK, et al. Inhibition of SARS-CoV 3C-like Protease Activity by Theaflavin-3,3'-digallate (TF3). Evid Based Complement Alternat Med 2005; 2: 209-15.
- [62] Lin CW, Tsai FJ, Tsai CH, et al. Anti-SARS coronavirus 3C-like protease effects of *Isatis indigotica* root and plant-derived phenolic compounds. Antiviral Res 2005; 68: 36-42.
- [63] Thiel V, Ivanov KA, Putics A, et al. Mechanisms and enzymes involved in SARS coronavirus genome expression. J Gen Virol 2003; 84: 2305-15.
- [64] Chen XP, Li GH, Tang XP, Xiong Y, Chen XJ, Cao Y. Lack of severe acute respiratory syndrome in 19 AIDS patients hospitalized together. J Acquir Immune Defic Syndr 2003; 34: 242-43.
- [65] Chen XP, Cao Y. Consideration of highly active antiretroviral therapy in the prevention and treatment of severe acute respiratory syndrome. Clin Infect Dis 2004; 38: 1030-32.
- [66] Chan KS, Lai ST, Chu CM, *et al.* Treatment of severe acute respiratory syndrome with lopinavir/ritonavir: a multicentre retrospective matched cohort study. Hong Kong Med J 2003; 9: 399-406.
- [67] Chu CM, Cheng VC, Hung IF, et al. Role of lopinavir/ritonavir in the treatment of SARS: initial virological and clinical findings. Thorax 2004; 59: 252-56.
- [68] Zhang XW, Yap YL. Old drugs as lead compounds for a new disease? Binding analysis of SARS coronavirus main proteinase with HIV, psychotic and parasite drugs. Bioorg Med Chem 2004; 12: 2517-21.
- [69] Chen F, Chan KH, Jiang Y, et al. in vitro Susceptibility of 10 clinical isolates of SARS coronavirus to selected antiviral compounds. J Clin Virol 2004; 31: 69-75.
- [70] Yamamoto N, Yang R, Yoshinaka Y, et al. HIV protease inhibitor nelfinavir inhibits replication of SARS-associated coronavirus. Biochem Biophys Res Commun 2004; 318: 719-25.
- [71] Xu X, Liu Y, Weiss S, Arnold E, Sarafianos SG, Ding J. Molecular model of SARS coronavirus polymerase: implications for biochemical functions and drug design. Nucleic Acids Res 2003; 31: 7117-30.
- [72] Parker WB. Metabolism and antiviral activity of ribavirin. Virus Res 2005; 107: 165-71.
- [73] Thomas HC, Torok ME, Forton DM, Taylor-Robinson SD. Possible mechanisms of action and reasons for failure of antiviral therapy in chronic hepatitis C. J Hepatol 1999; 31 Suppl 1: 152-59.
- [74] Tan EL, Ooi EE, Lin CY, et al. Inhibition of SARS coronavirus infection in vitro with clinically approved antiviral drugs. Emerg Infect Dis 2004; 10: 581-86.

Recent Patents on Anti-Infective Drug Discovery, 2006, Vol. 1, No. 3 307

- [75] Avendano M, Derkach P, Swan S. Clinical course and management of SARS in health care workers in Toronto: a case series. CMAJ 2003; 168: 1649-60.
- [76] van Vonderen MG, Bos JC, Prins JM, Wertheim-van Dillen P, Speelman P. Ribavirin in the treatment of severe acute respiratory syndrome (SARS). Neth J Med 2003; 61: 238-41.
- [77] Knowles SR, Phillips EJ, Dresser L, Matukas L. Common adverse events associated with the use of ribavirin for severe acute respiratory syndrome in Canada. Clin Infect Dis 2003; 37: 1139-42.
- [78] Barnard DL, Day CW, Bailey K, et al. Enhancement of the infectivity of SARS-CoV in BALB/c mice by IMP dehydrogenase inhibitors, including ribavirin. Antiviral Res 2006; 7: 53-63.
- [79] Barnard DL, Hubbard VD, Burton J, et al. Inhibition of severe acute respiratory syndrome-associated coronavirus (SARSCoV) by calpain inhibitors and beta-D-N4-hydroxycytidine. Antivir Chem Chemother 2004; 15: 15-22.
- [80] Hernandez-Santiago BI, Beltran T, Stuyver L, Chu CK, Schinazi RF. Metabolism of the anti-hepatitis C virus nucleoside beta-D-N4-hydroxycytidine in different liver cells. Antimicrob Agents Chemother 2004; 48: 4636-42.
- [81] Hoffmann P, Quasdorff M, Gonzalez-Carmona MA, Caselmann WH. Recent patents on experimental therapy for hepatitis C virus infection (1999 - 2002). Expert Opin Ther Pat 2003; 13: 1707-23.
- [82] Stuyver LJ, Whitaker T, McBrayer TR, et al. Ribonucleoside analogue that blocks replication of bovine viral diarrhea and hepatitis C viruses in culture. Antimicrob Agents Chemother 2003; 47: 244-54.
- [83] He R, Adonov A, Traykova-Adonova M, et al. Potent and selective inhibition of SARS coronavirus replication by aurintricarboxylic acid. Biochem Biophys Res Commun 2004; 320: 1199-203.
- [84] Yap Y, Zhang X, Andonov A, He R. Structural analysis of inhibition mechanisms of aurintricarboxylic acid on SARS-CoV polymerase and other proteins. Comput Biol Chem 2005; 29: 212-19.
- [85] De Clercq E. New inhibitors of human cytomegalovirus (HCMV) on the horizon. J Antimicrob Chemother 2003; 51: 1079-83.
- [86] Bisht H, Roberts A, Vogel L, Subbarao K, Moss B. Neutralizing antibody and protective immunity to SARS coronavirus infection of mice induced by a soluble recombinant polypeptide containing an N-terminal segment of the spike glycoprotein. Virology 2005; 334: 160-65.
- [87] Borowski P, Schalinski S, Schmitz H. Nucleotide triphosphatase/ helicase of hepatitis C virus as a target for antiviral therapy. Antiviral Res 2002; 55: 397-412.
- [88] Tanner JA, Zheng BJ, Zhou J, et al. The adamantane-derived bananins are potent inhibitors of the helicase activities and replication of SARS coronavirus. Chem Biol 2005; 12: 303-11.
- [89] Spiegel M, Pichlmair A, Martinez-Sobrido L, et al. Inhibition of Beta interferon induction by severe acute respiratory syndrome coronavirus suggests a two-step model for activation of interferon regulatory factor 3. J Virol 2005; 79: 2079-86.
- [90] Cheung CY, Poon LL, Ng IH, et al. Cytokine responses in severe acute respiratory syndrome coronavirus-infected macrophages in vitro: possible relevance to pathogenesis. J Virol 2005; 79: 7819-26.
- [91] Law HK, Cheung CY, Ng HY, et al. Chemokine upregulation in SARS coronavirus infected human monocyte derived dendritic cells. Blood 2005; 106: 2366-74.
- [92] Zheng B, He ML, Wong KL, et al. Potent inhibition of SARSassociated coronavirus (SCOV) infection and replication by type I interferons (IFN-alpha/beta) but not by type II interferon (IFNgamma). J Interferon Cytokine Res 2004; 24: 388-90.
- [93] Chen L, Liu P, Gao H, et al. Inhalation of nitric oxide in the treatment of severe acute respiratory syndrome: a rescue trial in Beijing. Clin Infect Dis 2004; 39: 1531-35.
- *[94] Carter, W.A., Strayer, D.: US20060035859 (2006).
- [95] Haagmans BL, Kuiken T, Martina BE, et al. Pegylated interferon-alpha protects type 1 pneumocytes against SARS coronavirus infection in macaques. Nat Med 2004; 10: 290-93.
- [96] Cinatl J, Morgenstern B, Bauer G, Chandra P, Rabenau H, Doerr HW. Treatment of SARS with human interferons. Lancet 2003; 362: 293-94.

- [97] Gao ZC, Zhu JH, Sun Y, et al. Clinical investigation of outbreak of nosocomial severe acute respiratory syndrome. Zhongguo Wei Zhong Bing Ji Jiu Yi Xue 2003; 15: 332-35.
- [98] Loutfy MR, Blatt LM, Siminovitch KA, et al. Interferon alfacon-1 plus corticosteroids in severe acute respiratory syndrome: a preliminary study. Jama 2003; 290: 3222-28.
- [99] Wu W, Wang J, Liu P, et al. A hospital outbreak of severe acute respiratory syndrome in Guangzhou, China. Chin Med J (Engl) 2003; 116: 811-18.
- [100] Zhao Z, Zhang F, Xu M, et al. Description and clinical treatment of an early outbreak of severe acute respiratory syndrome (SARS) in Guangzhou, PR China. J Med Microbiol 2003; 52: 715-20.
- [101] Bao M, Zhang Y, Wan M, et al. Anti-SARS-CoV immunity induced by a novel CpG oligodeoxynucleotide. Clin Immunol 2005; 118: 180-87.
- [102] De Clercq E. Interferon: ten stories in one. A short review of some of the highlights in the history of an almost quinquagenarian. Acta Microbiol Immunol Hung 2005; 52: 273-89.
- [103] Padalko E, Nuyens D, De Palma A, et al. The interferon inducer ampligen [poly(I)-poly(C12U)] markedly protects mice against coxsackie B3 virus-induced myocarditis. Antimicrob Agents Chemother 2004; 48: 267-74.
- [104] Leyssen P, Drosten C, Paning M, et al. Interferons, interferon inducers, and interferon-ribavirin in treatment of flavivirusinduced encephalitis in mice. Antimicrob Agents Chemother 2003; 47: 777-82.
- [105] Gowen BB, Barnard DL, Smee DF, et al. Interferon alfacon-1 protects hamsters from lethal pichinde virus infection. Antimicrob Agents Chemother 2005; 49: 2378-86.
- [106] Essey RJ, McDougall BR, Robinson WE Jr. Mismatched doublestranded RNA (polyI-polyC(12)U) is synergistic with multiple anti-HIV drugs and is active against drug-sensitive and drugresistant HIV-1 in vitro. Antiviral Res 2001; 51: 189-202.
- [107] Thompson KA, Strayer DR, Salvato PD, et al. Results of a double-blind placebo-controlled study of the double-stranded RNA drug polyI:polyC12U in the treatment of HIV infection. Eur J Clin Microbiol Infect Dis 1996; 15: 580-87.
- [108] Zhang MY, Choudhry V, Xiao X, Dimitrov DS. Human monoclonal antibodies to the S glycoprotein and related proteins as potential therapeutics for SARS. Curr Opin Mol Ther 2005; 7: 151-56.
- [109] Johnson S, Oliver C, Prince GA, et al. Development of a humanized monoclonal antibody (MEDI-493) with potent in vitro and in vivo activity against respiratory syncytial virus. J Infect Dis 1997; 176: 1215-24.
- [110] ter Meulen J, Bakker AB, van den Brink EN, et al. Human monoclonal antibody as prophylaxis for SARS coronavirus infection in ferrets. Lancet 2004; 363: 2139-41.
- [111] Wong VW, Dai D, Wu AK, Sung JJ. Treatment of severe acute respiratory syndrome with convalescent plasma. Hong Kong Med J 2003; 9: 199-201.
- [112] Traggiai E, Becker S, Subbarao K, et al. An efficient method to make human monoclonal antibodies from memory B cells: potent neutralization of SARS coronavirus. Nat Med 2004; 10: 871-75.

- [113] Yeh KM, Chiueh TS, Siu LK, *et al.* Experience of using convalescent plasma for severe acute respiratory syndrome among healthcare workers in a Taiwan hospital. J Antimicrob Chemother 2005; 56: 919-22.
- [114] Sui J, Li W, Murakami A, *et al*. Potent neutralization of severe acute respiratory syndrome (SARS) coronavirus by a human mAb to S1 protein that blocks receptor association. Proc Natl Acad Sci USA 2004; 101: 2536-41.
- [115] Sui J, Li W, Roberts A, *et al*. Evaluation of human monoclonal antibody 80R for immunoprophylaxis of severe acute respiratory syndrome by an animal study, epitope mapping, and analysis of spike variants. J Virol 2005; 79: 5900-06.
- [116] Ip WK, Chan KH, Law HK, et al. Mannose-binding lectin in severe acute respiratory syndrome coronavirus infection. J Infect Dis 2005; 191: 1697-704.
- [117] Wu CJ, Jan JT, Chen CM, et al. Inhibition of severe acute respiratory syndrome coronavirus replication by niclosamide. Antimicrob Agents Chemother 2004; 48: 2693-96.
- [118] Cinatl J, Morgenstern B, Bauer G, Chandra P, Rabenau H, Doerr HW. Glycyrrhizin, an active component of liquorice roots, and replication of SARS-associated coronavirus. Lancet 2003; 361: 2045-46.
- [119] Hoever G, Baltina L, Michaelis M, et al. Antiviral activity of glycyrrhizic acid derivatives against SARS-coronavirus. J Med Chem 2005; 48: 1256-59.
- [120] Harada S. Broad anti-viral agent glycyrrhizin directly modulates the fluidity of plasma membrane and HIV-1 envelope. Biochem J 2005; 392: 191-99.
- [121] Li B-j, Tang Q, Cheng D, et al. Using siRNA in prophylactic and therapeutic regimens against SARS coronavirus in *Rhesus* macaque. Nat Med 2005; 11: 944-51.
- [122] Li T, Zhang Y, Fu L, et al. siRNA targeting the leader sequence of SARS-CoV inhibits virus replication. Gene Ther 2005; 12: 751-61.
- [123] Tao P, Zhang J, Tang N, Zhang BQ, He TC, Huang AL. Potent and specific inhibition of SARS-CoV antigen expression by RNA interference. Chin Med J (Engl) 2005; 118: 714-19.
- [124] Shi Y, Yang de H, Xiong J, Jia J, Huang B, Jin YX. Inhibition of genes expression of SARS coronavirus by synthetic small interfering RNAs. Cell Res 2005; 15: 193-200.
- [125] Wu CJ, Huang HW, Liu CY, Hong CF, Chan YL. Inhibition of SARS-CoV replication by siRNA. Antiviral Res 2005; 65: 45-48.
- [126] Zhang R, Guo Z, Lu J, *et al*. Inhibiting severe acute respiratory syndrome-associated coronavirus by small interfering RNA. Chin Med J (Engl) 2003; 116: 1262-64.
- [127] He ML, Zheng BJ, Chen Y, et al. Kinetics and synergistic effects of siRNAs targeting structural and replicase genes of SARSassociated coronavirus. FEBS Lett 2006; 580: 2414-20.
- [128] Schubert S, Kurreck J. Oligonucleotide-based antiviral strategies. Handb Exp Pharmacol 2006; 261-87.
- *[129] Iversen, P.L., Stein, D.A.: US20060063150 (2006).
- [130] Song E, Zhu P, Lee S-K, et al. Antibody mediated in vivo delivery of small interfering RNAs via cell-surface receptors. Nat Biotech 2005; 23: 709-17.
- [131] Williams BR. Targeting specific cell types with silencing RNA. N Engl J Med 2005; 353: 1410-11.