

Short Communication

Detection of the Human Coronavirus 229E, HKU1, NL63, and OC43 between 2010 and 2013 in Yamagata, Japan

Yohei Matoba^{1*}, Chieko Abiko¹, Tatsuya Ikeda¹, Yoko Aoki¹, Yu Suzuki¹, Kazue Yahagi¹, Yoko Matsuzaki², Tsutomu Itagaki³, Fumio Katsushima⁴, Yuriko Katsushima⁴, and Katsumi Mizuta¹

¹Department of Microbiology, Yamagata Prefectural Institute of Public Health, Yamagata 990-0031;

²Department of Infectious Diseases, Yamagata University Faculty of Medicine, Yamagata 990-9585;

³Yamanobe Pediatric Clinic, Yamagata 990-0301; and

⁴Katsushima Pediatric Clinic, Yamagata 990-2461, Japan

SUMMARY: The available literature on human coronaviruses (HCoVs) in Japan is limited to epidemiological studies conducted over a maximum of 1 year. We conducted a 4-year study of HCoVs by analyzing 4,342 respiratory specimens obtained in Yamagata, Japan, between January 2010 and December 2013. A pan-coronavirus reverse transcription-PCR screening assay was performed, and all HCoV-positive specimens were subsequently confirmed by sequencing of the PCR products. We detected in 332 (7.6%) HCoV strains during the study period, comprising 133 (3.1%) HCoV-NL63, 83 (1.9%) HCoV-HKU1, 78 (1.8%) HCoV-OC43, and 38 (0.9%) HCoV-229E strains. HCoV detection per year ranged from 3.5% to 9.7%. HCoVs were detected mainly in winter, with January (28.5%) and February (25.3%) 2011 and December 2012 (14.6%) being the only months in which HCoV-NL63 detection per month exceeded 10.0%. HCoV-HKU1 displayed clear biennial peaks in January (18.3%) and February (10.7%) 2010 and in February (18.8%) and March (14.7%) 2012. The peak detection of HCoV-OC43 was 13.6% in November 2010, while that of HCoV-229E was 10.8% in March 2013. Our results indicated that there may be annual variations in the circulation of individual HCoV strains. Further long-term surveillance is necessary to clarify HCoV prevalence and circulation patterns in Japan.

Human coronaviruses (HCoVs) belong to the genus *Coronavirus* in the family *Coronaviridae* and include HCoV-229E, HCoV-OC43, and severe acute respiratory syndrome (SARS)-CoV (1). Recently, new HCoVs, such as HCoV-NL63, HCoV-HKU1, and Middle East respiratory syndrome (MERS)-CoV, have also been described (1). Four HCoVs (HCoV-229E, HCoV-OC43, HCoV-NL63, and HCoV-HKU1) are associated with a wide range of respiratory illness, including common colds, and with high morbidity outcomes, such as pneumonia and bronchiolitis (1–3). These four HCoVs are found globally, although the frequency of detection of these HCoVs varies with geographical location (1–20). Poor growth and a lack of cytopathic effects on cell cultures have been major deterrents to HCoV research in the past (1). However, with the development of polymerase chain reaction (PCR) technology, there has been broad and rapid development in the field of coronavirology (1).

In Japan, according to the National Epidemiological Surveillance of Infectious Diseases (NESID) system, only 306 HCoV-positive cases were reported in 19 prefectures between 2010 and 2013 (21), which accounted for only 0.4% of the total number of reported cases of

respiratory virus infection (306/68,332). Longitudinal studies of HCoV epidemiology are particularly lacking in Japan, with the available literature limited to descriptions representing a maximum of 1 year (4–7). Although we have conducted epidemiological surveys of acute viral respiratory infections based on virus isolation, we failed to isolate HCoVs in Yamagata. Thus, we performed a screening assay targeting HCoVs using reverse transcription-PCR (RT-PCR) to clarify the epidemiology of these viruses in Yamagata, Japan.

Between January 2010 and December 2013, 4,342 throat and nasal swab specimens were collected from patients with upper or lower acute respiratory infections at pediatric clinics in collaboration with the Yamagata Prefectural Institute of Public Health for the NESID. Specimens were transported to the Department of Microbiology at the Yamagata Prefectural Institute of Public Health for virus isolation. Among these specimens, 3,092 (71.2%) were from patients aged ≤ 5 years, 767 (17.7%) from patients aged 6–10 years, 326 (7.5%) from patients aged 11–15 years, 104 (2.4%) from patients aged > 15 years, and 53 (1.2%) from patients of unknown age. We were able to isolate several respiratory viruses, including influenza virus, parainfluenza virus, respiratory syncytial virus, human metapneumovirus, adenovirus, enterovirus, rhinovirus, parechovirus, mumps virus, cytomegalovirus, and herpes simplex virus, from 2,355 specimens using a microplate method with the following cell lines: HEF, HEp-2, VeroE6, MDCK, RD18s, GMK, HMV-II, and LLC-MK2 (22–24). We next investigated the presence of HCoV in the above specimens by RT-PCR analysis.

Viral RNA was extracted from 200 μ L of each speci-

Received June 18, 2014. Accepted July 22, 2014. J-STAGE Advance Publication November 25, 2014.

DOI: 10.7883/yoken.JJID.2014.266

*Corresponding author: Mailing address: Department of Microbiology, Yamagata Prefectural Institute of Public Health, Tokamachi 1-6-6, Yamagata, Yamagata 990-0031, Japan. Tel: +81-23-627-1373, Fax: +81-23-641-7486, E-mail: matobay@pref.yamagata.jp

men using the High Pure Virus RNA Kit (Roche Diagnostics, Mannheim, Germany) and was then transcribed into cDNA using the Prime Script™ RT Regent Kit (Takara Bio, Shiga, Japan), according to manufacturer's instructions. We next screened for the amplification of the four HCoVs (HCoV-229E, HCoV-OC43, HCoV-NL63, and HCoV-HKU1) by nested PCR using the PanCoVfw, PanCoVfw2, PanCoVr, and PanCoVr2 primers, which target the polymerase gene specific to the HCoV family, as reported previously (2). However, in our experience, human genomes are sometimes detected in amplification products using these primers. We found that such human genome amplification products included TTTAAA, which is not included in the amplification products of HCoVs. We, therefore, digested the amplification products using the restriction enzyme *DraI*, which recognizes TTTAAA. Using this step, we eventually succeeded in differentiating HCoV from human genomes. All HCoV-positive amplification products were then sequenced, and the four HCoVs were identified.

Figure 1 shows the monthly number and frequency of HCoVs detected. This report is the first to be based on a 4-year, long-term epidemiological study of HCoVs in Japan. We detected 332 (7.6%) HCoV strains during the study period, comprising 133 (3.1%) HCoV-NL63, 83 (1.9%) HCoV-HKU1, 78 (1.8%) HCoV-OC43, and 38 (0.9%) HCoV-229E strains. Our results also indicated that the frequency of HCoV detection per year

ranged from 3.5% in 2013 to 9.7% in 2012. The detection frequencies of HCoVs in respiratory samples reported in the literature are highly variable, ranging from 1.6% to 16.0%, depending on the country, study period, study population, and detection modality (2–20). The most commonly observed HCoV in our survey was HCoV-NL63, whereas many previous studies reported a predominance of HCoV-OC43 (2,5–15). Generally, HCoVs display high detection rates in winter months, with little to no detection in summer months in temperate regions (2,3,8–10,14–16). We also mainly detected HCoVs in winter in Yamagata, Japan, located in a northern temperate region.

Our results showed an annual detection frequency of 0.6–6.4% for HCoV-NL63. A monthly detection frequency exceeding 10% was observed for NCoV-NL63 in January (28.5%) and February (25.3%) 2011 and in December 2012 (14.6%). However, HCoV-NL63 was detected almost throughout the entire period from July 2011 to March 2013. HCoV-NL63 was also detected in Niigata from December 2010 to January 2011 (6). Combining data of HCoV detection in Yamagata with those in Niigata suggested that HCoV-NL63 circulated in the winter 2010/2011 season in the Niigata–Yamagata area.

Our results also indicated that the annual detection frequency of HKU1 was 0.1–3.4%. HCoV-HKU1 displayed clear biennial peaks from January (18.3%) to February (10.7%) in 2010 and February (18.8%) to March (14.7%) in 2012. HCoV-HKU1 peaked primarily

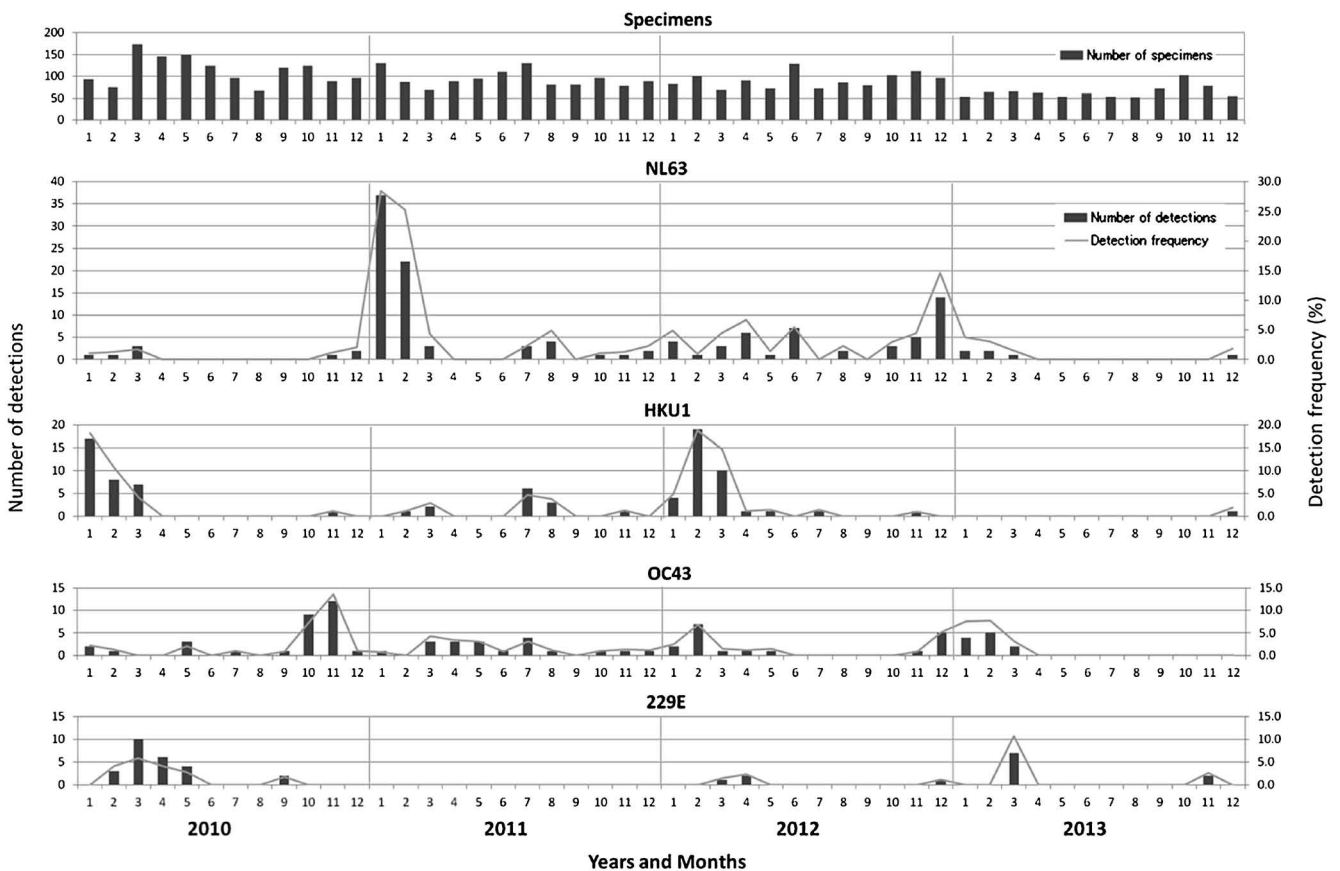


Fig. 1. Monthly distribution of human coronaviruses detected from patients with acute respiratory infections in Yamagata, Japan between 2010 and 2013. Bar indicates the number of detections. Detection frequency (%) per month is shown by the line.

Table 1. Coinfection of respiratory specimens with human coronaviruses and other viruses between 2010 and 2013 in Yamagata, Japan

Coinfecting isolated virus	HCoV-NL63 (n = 133)	HCoV-HKU1 (n = 83)	HCoV-OC43 (n = 78)	HCoV-229E (n = 38)	All HCoV (n = 332)
Enterovirus	8 (6.0)	2 (2.4)	2 (2.6)	2 (5.3)	14 (4.2)
HPIV	7 (5.3)	0	2 (2.6)	3 (7.9)	12 (3.6)
Adenovirus	4 (3.0)	2 (2.4)	4 (5.1)	1 (2.6)	11 (3.3)
Rhinovirus	7 (5.3)	2 (2.4)	1 (1.3)	0	10 (3.0)
HMPV	2 (1.5)	4 (4.8)	0	3 (7.9)	9 (2.7)
Influenza virus	3 (2.3)	2 (2.4)	0	0	5 (1.5)
RSV	1 (0.8)	1 (1.2)	1 (1.3)	1 (2.6)	4 (1.2)
Parechovirus	0	0	1 (1.3)	0	1 (0.3)
Mumps virus	0	1 (1.2)	0	0	1 (0.3)
CMV	6 (4.5)	1 (1.2)	3 (3.8)	0	10 (3.0)
HSV	0	1 (1.2)	0	0	1 (0.3)
HPIV, Rhinovirus	0	0	1 (1.3)	0	1 (0.3)
HPIV, Parechovirus	0	0	1 (1.3)	0	1 (0.3)
HMPV, CMV	0	0	0	1 (2.6)	1 (0.3)
Total	38 (28.6)	16 (19.3)	16 (20.5)	11 (28.9)	81 (24.4)

Data are no. (%) of patients.

HPIV, human parainfluenza virus; HMPV, human metapneumovirus; RSV, respiratory syncytial virus; CMV, cytomegalovirus; HSV, herpes simplex virus.

in winter and spring in 2010 and 2012, whereas it was detected only sporadically in summer and/or autumn in 2010, 2011, and 2012. This is the first report on HCoV-HKU1 circulation in Japan in the literature.

The annual detection frequency of HCoV-OC43, according to our data, was 1.4–2.2%. HCoV-OC43 detection showed a peak in November 2010 (13.6%) and was mainly detected from autumn to winter. However, HCoV-OC43 was detected almost throughout the period from May 2010 to May 2012. At the same time, HCoV-OC43 was also detected in Niigata from February to July 2011 (6). Our data of virus detection in Yamagata and Niigata suggest that HCoV-OC43 circulated in late winter, spring, and early summer in 2011 in the Niigata–Yamagata area. HCoV-OC43 was detected at a rate of 37.2% (29/78) in Mie from January to March 2013 (7). At the same time, the detection frequency was 7.5% in January and 7.8% in February in Yamagata. These data of virus detection in Yamagata and Mie suggest the possibility that HCoV-OC43 circulated in different areas of Japan from January to March 2013.

This study also showed that the annual detection frequency of HCoV-229E was 0–1.9%. HCoV-229E detection peaked in March 2013 (10.8%); however, it was detected at low levels throughout the study period. Many previous studies have also reported low frequencies of HCoV-229E compared with the frequencies of other HCoVs (2,3,5–11,14–17). HCoV-229E was mainly detected in spring, although the seasonal pattern of 229E could not be determined because of the small number of positive cases.

Our result and those of previous reports (3–10,12,14–16,19,20) suggest that HCoVs peak mainly in winter, although they can be detected throughout the year.

Table 1 shows respiratory specimens with coinfection of HCoVs and other viruses, which were identified by virus isolation using a microplate method between 2010 and 2013 in Yamagata, Japan. Of all 332 patients with

HCoV infections, 24.4% were coinfecting with other viruses. The respiratory viruses most frequently detected with HCoVs were enterovirus (4.2%), parainfluenza virus (3.6%), adenovirus (3.3%), and rhinovirus (3.0%). The coinfection rates of HCoVs in respiratory samples reported in the literature vary, ranging from 30.5% to 70.0% (2,3,12–20). Previous studies have demonstrated a high rate of HCoV coinfection with respiratory syncytial virus, influenza virus, and rhinovirus (2,3,13–19). However, as detection methods, PCR primers, and study designs often differ from one study to another, direct comparisons of data are difficult.

This study revealed that HCoVs were a significant cause of acute respiratory infections, accounting for 3.5–9.7% of such cases in Japan. Therefore, further long-term surveillance is necessary to more clearly evaluate the prevalence and circulation patterns of HCoVs in Japan.

Acknowledgments We thank the medical staff and people of Yamagata Prefecture for their collaboration in specimen collection for the NESID.

Conflict of interest None to declare.

REFERENCES

- McIntosh K, Englund JA. Coronaviruses and toroviruses, including severe acute respiratory syndrome. In: Cherry J, Harrison G, Kaplan S, et al. editors. Feigin and Cherry's Textbook of Pediatric Infectious Diseases. 7th ed. Philadelphia: Elsevier Saunders; 2014. p. 2486-95.
- Canducci F, Debiaggi M, Sampaolo M, et al. Two-year prospective study of single infections and co-infections by respiratory syncytial virus and viruses identified recently in infants with acute respiratory disease. *J Med Virol.* 2008;80:716-23.
- Jevšnik M, Uršič T, Zigon N, et al. Coronavirus infections in hospitalized pediatric patients with acute respiratory tract disease. *BMC Infect Dis.* 2012;12:365.
- Suzuki Y, Watanabe O, Okamoto M, et al. Detection of human coronavirus NL63 from children with respiratory illness in 2003–Sendai City. *Infect Agents Surveillance Rep.* 2004;25:181-2. Japanese.

5. Kaida A, Kubo H, Sekiguchi J, et al. Experiment of multiplex real-time PCR for detection of various respiratory viruses, October 2009–September 2010–Osaka City. *Infect Agents Surveillance Rep.* 2011;32:202-3. Japanese.
6. Kon M, Watanabe K, Tazawa T, et al. Detection of human coronavirus NL63 and OC43 in children with acute respiratory infections in Niigata, Japan, between 2010 and 2011. *Jpn J Infect Dis.* 2012;65:270-2.
7. Yano T, Maeda C, Kobayashi A, et al. Human coronavirus isolates from infants with respiratory symptoms (January–April 2013)–Mie Prefecture. *Infect Agents Surveillance Rep.* 2013;34:170-2. Japanese.
8. Talbot HK, Shepherd BE, Crowe JE Jr, et al. The pediatric burden of human coronaviruses evaluated for twenty years. *Pediatr Infect Dis J.* 2009;28:682-7.
9. Lau SK, Woo PC, Yip CC, et al. Coronavirus HKU1 and other coronavirus infections in Hong Kong. *J Clin Microbiol.* 2006;44:2063-71.
10. Woo PC, Yuen KY, Lau SK. Epidemiology of coronavirus-associated respiratory tract infections and the role of rapid diagnostic tests: a prospective study. *Hong Kong Med J.* 2012;18:S22-4.
11. Dijkman R, Jebbink MF, Gaunt E, et al. The dominance of human coronavirus OC43 and NL63 infections in infants. *J Clin Virol.* 2012;53:135-9.
12. Kim JK, Jeon JS, Kim JW, et al. Epidemiology of respiratory viral infection using multiplex RT-PCR in Cheonan, Korea (2006–2010). *J Microbiol Biotechnol.* 2013;23:267-73.
13. Dare RK, Fry AM, Chittaganpitch M, et al. Human coronavirus infections in rural Thailand: a comprehensive study using real-time reverse-transcription polymerase chain reaction assays. *J Infect Dis.* 2007;196:1321-8.
14. Gaunt ER, Hardie A, Claas EC, et al. Epidemiology and clinical presentations of the four human coronaviruses 229E, HKU1, NL63, and OC43 detected over 3 years using a novel multiplex real-time PCR method. *J Clin Microbiol.* 2010;48:2940-7.
15. Prill MM, Iwane MK, Edwards KM, et al. Human coronavirus in young children hospitalized for acute respiratory illness and asymptomatic controls. *Pediatr Infect Dis J.* 2012;31:235-40.
16. Kuypers J, Martin ET, Heugel J, et al. Clinical disease in children associated with newly described coronavirus subtypes. *Pediatrics.* 2007;119:e70-6.
17. Talbot HK, Crowe JE Jr, Edwards KM, et al. Coronavirus infection and hospitalizations for acute respiratory illness in young children. *J Med Virol.* 2009;81:853-6.
18. Cabeça TK, Granato C, Bellei N. Epidemiological and clinical features of human coronavirus infections among different subsets of patients. *Influenza Other Respir Viruses.* 2013;7:1040-7.
19. Lu R, Yu X, Wang W, et al. Characterization of human coronavirus etiology in Chinese adults with acute upper respiratory tract infection by real-time RT-PCR assays. *PLoS One.* 2012;7:e38638.
20. Lepiller Q, Barth H, Lefebvre F, et al. High incidence but low burden of coronaviruses and preferential associations between respiratory viruses. *J Clin Microbiol.* 2013;51:3039-46.
21. National Epidemiological Surveillance of Infectious Diseases (NESID) system. Available at <<https://nesid3g.wish.mhlw.hq.admix.go.jp/>>. Accessed May 8, 2014.
22. Mizuta K, Abiko C, Aoki Y, et al. Analysis of monthly isolation of respiratory viruses from children by cell culture using a microplate method: a two-year study from 2004 to 2005 in Yamagata, Japan. *Jpn J Infect Dis.* 2008;61:196-201.
23. Mizuta K, Abiko C, Aoki Y, et al. Epidemiology of parainfluenza virus types 1, 2 and 3 infections based on virus isolation between 2002 and 2011 in Yamagata, Japan. *Microbiol Immunol.* 2012;56:822-8.
24. Abiko C, Mizuta K, Aoki Y, et al. An outbreak of parainfluenza virus type 4 infections among children with acute respiratory infections during the 2011–2012 winter season in Yamagata, Japan. *Jpn J Infect Dis.* 2013;66:76-8.