

## 222 Human Coronaviruses

Susan M. Poutanen

Coronaviruses are included in the Coronaviridae family under the order Nidovirales. They are enveloped, nonsegmented, single-stranded, positive-sense RNA viruses named after their corona-like or crown-like surface projections seen on electron microscopy that correspond to large surface spike proteins (Figs. 222.1 and 222.2). They are host specific and can infect humans and a variety of animals.<sup>1</sup>

Four genera of coronaviruses have been described. Human coronaviruses (HCoVs) are part of the *Alphacoronavirus* and *Betacoronavirus* genera<sup>1,2</sup> (Table 222.1).

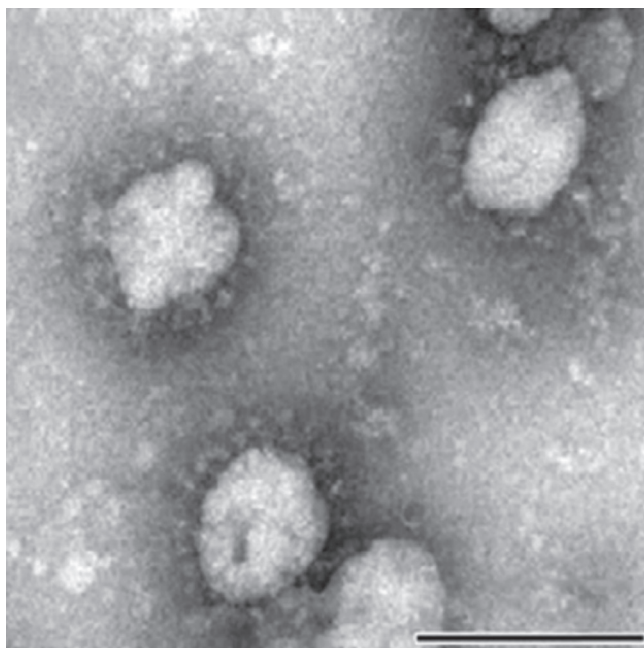
### EPIDEMIOLOGY

In the 1930s, coronaviruses were recognized as animal pathogens.<sup>3</sup> Thirty years later, coronaviruses were identified as human respiratory

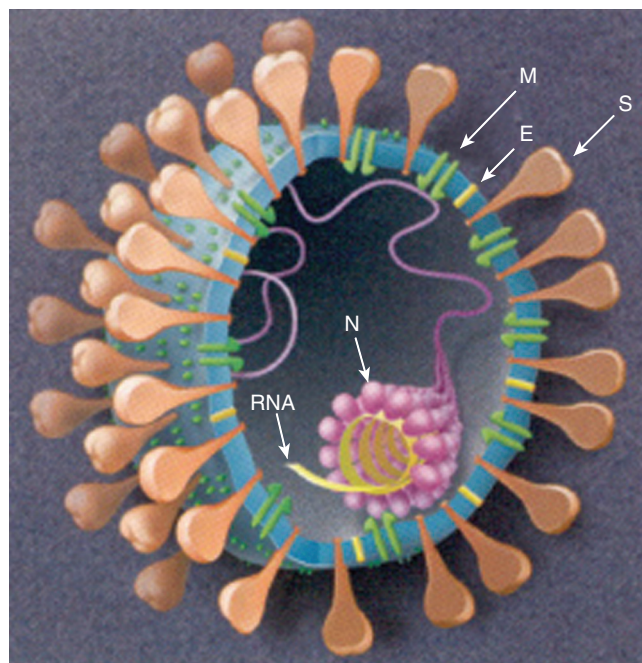
pathogens. The first recognized HCoV strains included 229E and OC43. Strains such as B814, OC16, OC37, and OC48 also were described but were not characterized further.<sup>4-6</sup> Coronavirus-like particles have been detected in stool, primarily in infants with gastroenteritis and necrotizing enterocolitis, but further characterization has not been completed.<sup>7-9</sup>

In 2003, severe acute respiratory syndrome (SARS) CoV was identified as a novel respiratory pathogen responsible for a global outbreak of SARS. First emerging in 2002 in China, the outbreak lasted 9 months and resulted in 8098 people infected and 774 deaths.<sup>10-14</sup> Data suggest that SARS CoV evolved from SARS CoV-like viruses in horseshoe bats, with civet cats and other wild market animals serving as intermediate hosts.<sup>15-20</sup>

Renewed interest in CoV research led to the discovery of two novel HCoVs 2 years later: NL63 (also known as NL or NH) and HKU1.<sup>21-23</sup>



**FIGURE 222.1** Negative-contrast electron micrograph of severe acute respiratory syndrome coronaviruses shows the typical crown-like spike proteins on the surface of the coronavirus particles (bar = 100 nm). (From Kuiken T, Fouchier RA, Schutten M, et al. Newly discovered coronavirus as the primary cause of severe acute respiratory syndrome. *Lancet* 2003;362:263–270.)



**FIGURE 222.2** Organization of the spike (S), membrane (M), and envelope (E) glycoproteins in a typical coronavirus is shown for a typical coronavirus. The RNA is protected by the nucleocapsid proteins (N). (From Holmes KV, Enjuanes L. The SARS coronavirus: a postgenomic era. *Science* 2003;300:1377–1378.)

HCoV-NL63 has been detected in human respiratory samples from as early as 1981.<sup>24</sup> It is unclear how HCoV-NL63 and HCoV-HKU1 relate to the HCoV strains originally described in the 1960s (i.e., B814, OC16, OC37, and OC48) or to the enteric coronavirus-like particles detected in stool.<sup>25</sup>

In 2012, the Middle East respiratory syndrome (MERS) CoV (also called hCoV-EMC) was identified as a novel CoV responsible for an epidemic of respiratory illness in the Kingdom of Saudi Arabia.<sup>26,27</sup> Data suggest that MERS CoV likely evolved from bat CoV, with camels acting as intermediate hosts.<sup>28</sup> A total of 1733 cases and 678 deaths were associated with MERS CoV as of May 2016.<sup>29</sup>

HCoV-229E, HCoV-OC43, HCoV-NL63, and HCoV-HKU1 are found worldwide and cause disease predominantly in winter and spring months in temperate climates.<sup>24,30</sup> Seroprevalence data suggest that exposure is common in early childhood.<sup>31,32</sup> SARS CoV has not been identified since 2004, when 4 sporadic community-acquired cases of SARS were identified

in China<sup>33</sup> and 13 cases linked to laboratory biosafety practice breaches were identified in Southeast Asia.<sup>34–36</sup> During the 2002–2003 outbreak, most SARS CoV transmission occurred within hospitals from patients with unrecognized illness.<sup>37</sup> MERS CoV cases continue in the Middle East, primarily linked to exposure to camel or camel products or close contact with unrecognized cases in hospitalized patients.<sup>38–40</sup> Household transmission also has occurred.<sup>41</sup> In 2015, a large outbreak of MERS CoV in South Korea, affecting 186 cases and associated with 36 deaths, was traced to an individual who returned from travel in the Middle East.<sup>28,29,42</sup>

Modes of transmission for HCoV other than SARS CoV and MERS CoV have not been well studied. Based on studies of other respiratory viruses, transmission likely occurs primarily by a combination of spread by droplet and direct and indirect contact.<sup>43–45</sup> The possible role of aerosol spread needs further study. Droplet spread and direct contact likely are the most common modes of transmission for SARS CoV, although evidence for spread by indirect contact and aerosol also

**TABLE 222.1** Human Coronaviruses

Genus	Species	Acronym	Host	Associated Diseases	
<b>Alphacoronavirus</b>	Human CoV-229E	HCoV-229E	Human	Respiratory tract infection	
	Human CoV-NL63	HCoV-NL63	Human	Respiratory tract infection	
<b>Betacoronavirus</b>	Subgroup A	Human CoV-OC43 <sup>a</sup>	Human	Respiratory tract infection	
		Human CoV-HKU1	Human	Respiratory tract infection and possibly gastroenteritis	
	Subgroup B	SARS CoV	SARS CoV	Human	Respiratory tract infection
	Subgroup C	MERS CoV	MERS CoV	Human	Respiratory tract infection
	Subgroup D	No human CoV identified	—	—	—
<b>Deltacoronavirus</b>	No human CoV identified	—	—	—	
<b>Gammacoronavirus</b>	No human CoV identified	—	—	—	

<sup>a</sup>This species has been abolished according to the International Committee on Taxonomy of Viruses. It is now considered part of the species *betacoronavirus 1*.<sup>1</sup> However, because the name HCoV-OC43 is still commonly used, the species is referred to as HCoV-OC43 in this chapter.

CoV, coronavirus; HCoV, human coronavirus; MERS, Middle East respiratory syndrome; SARS, Severe acute respiratory syndrome.

exists.<sup>46–53</sup> There is no evidence of vertical transmission of SARS CoV.<sup>54,55</sup> Transmission of MERS CoV is thought to occur through droplet, contact, and possibly aerosol routes.<sup>40,41,56</sup>

HCoV-229E and HCoV-OC43 are most likely transmitted during the first few days of illness, when symptoms and respiratory tract viral load are maximal.<sup>57,58</sup> Further study is needed to confirm if this also is the case for HCoV-NL63 and HCoV-HKU1. SARS CoV and MERS CoV are most likely transmitted during the second week of illness, when symptoms and viral load in the respiratory tract peak.<sup>59–62</sup> Supershedding events have been associated with SARS CoV and MERS CoV.<sup>39</sup>

The incubation period for HCoV-229E is 2 to 5 days (median, 3 days).<sup>58,63</sup> Further study is needed to confirm the incubation periods for HCoV-OC43, HCoV-NL63, and HCoV-HKU1. The incubation period for SARS CoV is 2 to 10 days (median, 4 days).<sup>59</sup> The incubation for MERS CoV is 2 to 15 days (median, 5 days).<sup>40</sup>

## PATHOGENESIS AND IMMUNITY

The pathogenesis of HCoVs has been best described for HCoV-229E, SARS CoV, and MERS CoV. For SARS CoV, most evidence is from infections in adults because few children were affected by the 2002–2003 outbreak.<sup>64</sup>

**Human Coronavirus 229E.** HCoV-229E infections are initiated through inoculation of respiratory tract mucosal surfaces. Nasal mucosal plasma exudation and increased interferon  $\gamma$  (IFN $\gamma$ ) levels in nasal lavage specimens correlate with symptom severity.<sup>65,66</sup> Respiratory tract viral loads peak within the first 3 days after infection and drop off dramatically at 1 week, correlating with development and improvement in symptoms.<sup>57,67</sup> Antibodies can be detected at 1 week, correlating with the drop in viral load, and they reach maximal levels approximately 1 week later and decline thereafter.<sup>68</sup>

Immunity is not complete, and reinfection is common.<sup>68,69</sup> Higher circulating antibody levels, especially levels of specific IgA anti-HCoV, correlate with reduced symptoms and reduced virus shedding on re-exposure.<sup>68,70</sup>

**Severe Acute Respiratory Syndrome Coronavirus.** SARS CoV infection is initiated through inoculation of the respiratory tract mucosa using angiotensin-converting enzyme 2 acting as the functional receptor for cell entry.<sup>71</sup> Viremia and replication in the lung and gastrointestinal tract follows.<sup>72,73</sup> Replication at other sites likely occurs given the wide distribution of SARS CoV in tissues examined at autopsy.<sup>74,75</sup>

Peak viral loads in nasopharyngeal specimens are detected during the second week of symptoms.<sup>60,73</sup> A rise in SARS CoV-specific antibodies typically is seen at the same time. Increasing antibody titers and symptomatic improvement during the second and third week are associated with a decrease in SARS CoV viral loads.<sup>60,76</sup> Paradoxically, despite a fall in viral load and a rise in SARS-specific antibodies, clinical deterioration is observed in some patients.<sup>60</sup> The host immune responses likely contribute to clinical deterioration in these patients. Elevated levels of IFN $\gamma$ , inflammatory cytokines interleukin-1 (IL-1), IL-6, and IL-12; neutrophil chemokine IL-8; monocyte chemoattractant protein 1; and IFN $\gamma$ -inducible protein-10 have been detected, with levels of IL-6 correlating with severity of disease.<sup>77,78</sup>

**Middle East Respiratory Syndrome Coronavirus.** MERS CoV infection most likely is initiated through inoculation of the respiratory tract mucosa mediated by dipeptidyl peptidase 4 (DPP4) (i.e., CD26) acting as the functional receptor.<sup>79,80</sup> Viral loads are highest in lower respiratory tract specimens, but real-time polymerase chain reaction (rt-PCR) evidence of MERS CoV also can be found in the upper respiratory tract, serum, stool, and urine.<sup>62</sup> Autopsy data from a single patient did not show extrapulmonary MERS CoV dissemination, but because DPP4 is widely distributed in different tissues, extrapulmonary dissemination is thought to be possible.<sup>79</sup>

## CLINICAL MANIFESTATIONS

**Human Coronaviruses 229E, OC43, NL63, and HKU1.** HCoVs 229E, OC43, NL63, and HKU1 are commonly associated with the common cold, which is typically characterized by rhinorrhea, nasal congestion, sore throat, sneezing, and cough that may be associated with fever.<sup>22,24,67,81–84</sup> Together, the HCoVs are the second most common cause of the common cold after rhinoviruses.<sup>85,86</sup>

Based on data for HCoV-229E, symptoms typically peak on day 3 or 4 of illness and are self-limited.<sup>67,87</sup> HCoVs also may be associated with acute otitis media or exacerbations of asthma.<sup>24,82,84,88,89</sup> Less frequently, these viruses are associated with lower respiratory tract infections, including bronchiolitis and pneumonia, primarily in infants and immunocompromised children and adults.<sup>23,82,84,90–98</sup>

Compared with other HCoVs, HCoV-NL63 more frequently is associated with croup and is the second most common cause after parainfluenza virus type 1.<sup>99,100</sup> A possible association of HCoV-NL63 with Kawasaki disease was not substantiated.<sup>101,102</sup>

HCoV-HKU1 has been associated with symptoms of gastroenteritis, including vomiting and diarrhea, which typically occur along with respiratory symptoms.<sup>84,89,103</sup> HCoV-HKU1 also appears to be more frequently associated with febrile seizures compared with other HCoVs.<sup>84,89</sup>

**Severe Acute Respiratory Syndrome Coronavirus.** SARS CoV is associated with severe symptoms.<sup>104–106</sup> SARS CoV disproportionately affects adults, who typically manifest fever, myalgia, headache, malaise, and chills, followed by a nonproductive cough and dyspnea 3 to 5 days later. Approximately 25% develop watery diarrhea. Respiratory distress progresses and requires intubation and ventilation in 25% of cases. The overall associated mortality rate is approximately 10%, with most deaths occurring in the third week of illness.<sup>105</sup> The case-fatality rate for persons older than 60 years of age approaches 50%.<sup>107</sup>

Typical laboratory abnormalities include lymphopenia and increased serum lactate dehydrogenase (LDH) and creatine kinase levels.<sup>108,109</sup> Most patients have progressive unilateral or bilateral, ill-defined airspace infiltrates on chest imaging.<sup>108,110–112</sup> Pneumothoraces and other signs of barotrauma are common in patients receiving mechanical ventilation.<sup>105</sup>

Infants and children younger than 12 years of age who develop SARS typically have fever, cough, rhinorrhea, and milder symptoms compared with adolescents and adults. Associated lymphopenia is less severe, and radiographic changes are milder and usually resolve more quickly than in adolescents and adults. No infants or children died of SARS CoV infection in the 2002–2003 outbreak.<sup>64,113–116</sup> Adolescents who developed SARS had clinical courses more closely resembling that of adults, including fever, myalgia, headache, and chills, and they were more likely to have dyspnea, hypoxemia, and worsening chest radiographic findings.

Women infected with SARS CoV during pregnancy who survive have an increased risk of spontaneous miscarriage, preterm delivery, and intrauterine growth restriction.<sup>54,55,117</sup> Two neonates born to mothers with SARS in the 2002–2003 outbreak developed gastrointestinal complications (e.g., jejunal perforation, necrotizing enterocolitis with ileal perforation) soon after birth, but neither had clinical evidence of SARS CoV infection.<sup>55</sup> It is unclear whether these findings related to complications of maternal SARS or treatments for SARS, such as ribavirin and corticosteroids, used during pregnancy.

**Middle East Respiratory Syndrome Coronavirus.** MERS CoV infection is associated with severe symptoms similar to those seen with SARS CoV, although a spectrum of disease, including asymptomatic infections and mild disease, can occur.<sup>27</sup> Most cases have been identified in male adults with comorbidities. Infected children typically have milder symptoms.<sup>27</sup> Fever, myalgia, and chills are followed a few days later by a nonproductive cough and dyspnea. Approximately 25% of patients also have vomiting, diarrhea, or abdominal pain.<sup>27</sup> Rapid deterioration of oxygenation with progressive unilateral or bilateral airspace infiltrates on chest imaging may follow, requiring mechanical ventilation and often associated with acute renal failure.<sup>118</sup>

Laboratory abnormalities include thrombocytopenia, lymphopenia, and an elevated LDH level.<sup>27</sup> The case-fatality rate is high, estimated at 36%.<sup>28</sup>

## DIAGNOSIS

Some laboratories offer comprehensive rt-PCR for respiratory tract specimens, which may include detection of HCoVs 229E, OC43, NL54, and HKU1.<sup>84,119</sup> Public health laboratories offer rt-PCR and antibody testing for SARS CoV and MERS CoV testing.<sup>120,121</sup>

Upper and lower respiratory tract specimens are the most appropriate samples for viral detection when testing is available.<sup>62,73,82,84,122</sup> Stool

samples frequently are positive for patients with SARS and have been positive for some with MERS CoV<sup>62</sup> and HCoV-HKU1 infection.<sup>73,89,103,122</sup> Serum samples also may be positive by rt-PCR for patients with SARS CoV and MERS CoV infection.<sup>62</sup> For cases of HCoV-229E and HCoV-OC43 infection, specimens are most likely to be positive during the first few days of illness<sup>57</sup>; whether this also is true for HCoV-NL63 and HCoV-HKU1 needs further study. For SARS CoV infection, serum samples for RT-PCR testing are most likely to be positive in the first week of illness,<sup>72,123</sup> but respiratory and stool specimens may not be positive until the second week of illness, when symptoms and viral loads peak.<sup>60,73</sup> Infants and children with SARS CoV infections are less likely to have positive specimens, consistent with the milder symptoms and presumed correspondingly lower viral loads in children.<sup>113,114</sup> For cases of MERS CoV infection, specimens for rt-PCR should be collected from the lower respiratory and upper respiratory tract along with serum and stool samples.<sup>62,124</sup>

Laboratory guidance for SARS CoV and MERS CoV diagnostic testing is available on the Centers for Disease Control and Prevention (CDC) website.<sup>124,125</sup> Because of the potential for false-positive results and the associated public health implications, testing for SARS CoV in the absence of known person-to-person transmission should be done only in consultation with public health departments and when there is a high degree of clinical suspicion. Similarly, testing for MERS CoV should be done only in consultation with public health officials and in the context of known risk factors for MERS CoV infection.

## TREATMENT

Because of the self-limited nature of infection with HCoV strains 229E, OC43, NL63, and HKU1, few treatment studies have been performed. Care typically is supportive.

SARS CoV infections are more serious. Corticosteroids, type 1 IFN agents, convalescent plasma, ribavirin, and lopinavir or ritonavir have been used to treat SARS.<sup>108,126–129</sup> For many of these agents, anecdotal reports suggest benefit, and in vitro assays and animal models offer supportive data,<sup>108,126–138</sup> with the exception of ribavirin, for which in vitro studies do not support efficacy.<sup>135,139,140</sup>

Since the SARS outbreak, viral entry- and protease-inhibiting agents, RNA-interfering agents, and glycyrrhizin have been tested in vitro and appear promising.<sup>140,141</sup> However, no definitive conclusions can be drawn. If SARS CoV re-emerges, clarification of the effectiveness of these treatments through controlled clinical trials will be needed. For MERS CoV, preliminary data suggest several treatments may be useful, but no definitive recommendations can be made at this time.<sup>142</sup>

## PREVENTION

Practicing good hand and respiratory hygiene is the most useful control measure to curb the spread of all respiratory viruses, including HCoVs.<sup>143,144</sup> Prophylactic intranasal IFN $\alpha$  has reduced the duration and severity of HCoV-229E infection in research settings, but it has not been used clinically.<sup>145,146</sup> A proprietary extract of the roots of North American ginseng (*Panax quinquefolium*) can reduce the number of colds and the severity and duration of cold symptoms in adults when taken daily, presumably due to immune stimulation.<sup>147–150</sup> Efficacy for a decrease in the number of colds specifically due to HCoVs has not been studied.

Healthcare personnel should use a gown, gloves, mask, and eye protection for the duration of illness when caring for children hospitalized with signs and symptoms of a respiratory tract infection.<sup>151</sup> The same precautions, with the replacement of the mask by a respirator if available, and negative-pressure isolation are recommended for patients with SARS CoV infection for the duration of illness or 10 days after resolution of fever, provided respiratory symptoms are absent or improving.<sup>151</sup> The same precautions recommended for SARS CoV are recommended for MERS CoV except the duration of precautions should be decided in conjunction with public health authorities.<sup>152</sup>

Standard disinfectants should be used to clean and disinfect environmental surfaces that are frequently touched by infected persons. This can decrease the potential for indirect transmission of HCoVs by fomites.<sup>153</sup>

The control of the 2002–2003 SARS outbreak is credited to the rapid identification of cases and early implementation of infection control and public health measures, including contact tracing and quarantine. If SARS CoV re-emerges, all measures should be

### Key Points: Epidemiology, Clinical Manifestations, Diagnosis, and Treatment of Human Coronavirus (HCoV) Infections

#### EPIDEMIOLOGY

- Human coronaviruses (HCoVs) 229E and OC43 (identified in the 1960s) and NL63 and HKU1 (identified in 2004) are found worldwide. Exposure is common in early childhood; in temperate climates, it primarily causes infections in the winter and spring months.
- Severe acute respiratory syndrome coronavirus (SARS CoV) was identified in 2003 as responsible for the 2002–2003 global outbreak of SARS.
- SARS CoV has not been associated with disease since 2004, and the possibility or probability of a large-scale re-emergence of SARS is unknown.
- Middle East respiratory syndrome coronavirus (MERS CoV) was identified in 2012 as responsible for an epidemic of respiratory illness in the Kingdom of Saudi Arabia. Transmission is ongoing and is primarily limited to countries in the Middle East.
- Risk factors include exposure to camels or camel products or close contact with persons infected with MERS CoV.
- The most common modes of transmission are through droplet and direct and indirect contact.

#### CLINICAL MANIFESTATIONS

- HCoV strains 229E, OC43, NL63, and HKU1 are associated with the common cold, acute otitis media, asthma exacerbations, and less frequently, bronchiolitis and pneumonia.
- HCoV-NL63 is also associated with croup. HCoV-HKU1 is associated with vomiting and diarrhea and frequently with

respiratory tract symptoms; it appears to be associated more frequently with febrile seizures compared with other HCoVs.

- SARS CoV causes SARS, with an attendant mortality rate of 10%, which primarily affects adults and adolescents. Children younger than 12 years of age who develop SARS typically have less severe manifestations (i.e., fever, cough, and rhinorrhea).
- MERS CoV is associated with a spectrum of disease, ranging from asymptomatic and mild disease to severe respiratory disease with an associated mortality rate of 36%.
- MERS CoV primarily affects male adults with comorbidities; children with MERS typically have milder symptoms.

#### DIAGNOSIS

- Upper and lower respiratory tract specimens can be tested by HCoV real-time polymerase chain reaction.
- Stool samples frequently are positive for patients with SARS CoV and have been positive for some patients with MERS CoV and HCoV-HKU1 infection; antibody tests also are available for SARS CoV and MERS CoV infection.
- Serum samples may be positive for patients infected with SARS CoV and MERS CoV.

#### TREATMENT

- Infections with HCoV strains 229E, OC43, NL63, and HKU1 require supportive care.
- SARS CoV and MERS CoV infections have no definitive treatment recommendations because of a lack of controlled trials.



implemented urgently in an attempt to prevent a recurrent worldwide outbreak.<sup>28,154,155</sup>

Transmission of MERS CoV within hospitals and households can be averted with the use of infection control precautions,<sup>55</sup> but preventing the transmission from camels to humans is more challenging given the prevalent use of camels for transport, meat, and milk. Recommendations to reduce the likelihood of transmission include regulation of camel movement, enforcing the use of personal protective precautions while handling camels, and educating the public about the risks of consuming unpasteurized camel milk and urine.<sup>156</sup> Given the challenges of following these recommendations, it is likely that sporadic transmission will continue until an effective MERS CoV vaccine is found.<sup>28,157</sup>

*All references are available online at [www.expertconsult.com](http://www.expertconsult.com).*

## KEY REFERENCES

10. Drosten C, Gunther S, Preiser W, et al. Identification of a novel coronavirus in patients with severe acute respiratory syndrome. *N Engl J Med* 2003;348:1967–1976.
11. Ksiazek TG, Erdman D, Goldsmith CS, et al. A novel coronavirus associated with severe acute respiratory syndrome. *N Engl J Med* 2003;348:1953–1966.
21. van der Hoek L, Pyrc K, Jebbink MF, et al. Identification of a new human coronavirus. *Nat Med* 2004;10:368–373.
22. Fouchier RA, Hartwig NG, Bestebroer TM, et al. A previously undescribed coronavirus associated with respiratory disease in humans. *Proc Natl Acad Sci USA* 2004;101:6212–6216.
23. Woo PC, Lau SK, Chu CM, et al. Characterization and complete genome sequence of a novel coronavirus, coronavirus HKU1, from patients with pneumonia. *J Virol* 2005;79:884–895.
26. Zaki AM, van Boheemen S, Bestebroer TM, et al. Isolation of a novel coronavirus from a man with pneumonia in Saudi Arabia. *N Engl J Med* 2012;367:1814–1820.
28. Hu B, Ge X, Wang LF, Shi Z. Bat origin of human coronaviruses. *Virol J* 2015; 12:221.
124. Centers for Disease Control and Prevention. CDC laboratory testing for Middle East respiratory syndrome coronavirus (MERS-CoV). <http://www.cdc.gov/coronavirus/mers/lab/lab-testing.html>.
125. Centers for Disease Control and Prevention. Public health guidance for community-level preparedness and response to severe acute respiratory syndrome (SARS), version 2/3. Appendix F8: guidelines for laboratory diagnosis of SARS-CoV infection. Supplement F: laboratory guidance. <http://www.cdc.gov/sars/guidance/f-lab/app8.html>.
151. Siegel JD, Rhinehart E, Jackson M, et al. Centers for Disease Control and Prevention: 2007 guideline for isolation precautions: preventing transmission of infectious agents in healthcare settings. <http://www.cdc.gov/hicpac/pdf/isolation/Isolation2007.pdf>.
152. Centers for Disease Control and Prevention. Interim infection prevention and control recommendations for hospitalized patients with Middle East respiratory syndrome coronavirus (MERS-CoV). <https://www.cdc.gov/coronavirus/mers/infection-prevention-control.html>.

## REFERENCES

- International Committee on Taxonomy of Viruses. Virus Taxonomy: 2015 Release. EC 47 London, UK, July 2015. Email ratification 2016 (MSL#30). <http://www.ictvonline.org/virusTaxonomy.asp>.
- Chan JF, Lau SK, To KK, et al. Middle East respiratory syndrome coronavirus: another zoonotic betacoronavirus causing SARS-like disease. *Clin Microbiol Rev* 2015;28:465–522.
- Kahn JS. The widening scope of coronaviruses. *Curr Opin Pediatr* 2006;18:42–47.
- McIntosh K. Coronaviruses in the limelight. *J Infect Dis* 2005;191:489–491.
- Tyrrell DA, Bynoe ML. Cultivation of a novel type of common-cold virus in organ cultures. *Br Med J* 1965;5448:1467–1470.
- McIntosh K, Dees JH, Becker WB, et al. Recovery in tracheal organ cultures of novel viruses from patients with respiratory disease. *Proc Natl Acad Sci USA* 1967;57:933–940.
- Mortensen ML, Ray CG, Payne CM, et al. Coronaviruslike particles in human gastrointestinal disease: epidemiologic, clinical, and laboratory observations. *Am J Dis Child* 1985;139:928–934.
- Resta S, Luby JP, Rosenfeld CR, et al. Isolation and propagation of a human enteric coronavirus. *Science* 1985;229:978–981.
- Gerna G, Passarini N, Battaglia M, et al. Human enteric coronaviruses: antigenic relatedness to human coronavirus OC43 and possible etiologic role in viral gastroenteritis. *J Infect Dis* 1985;151:796–803.
- Drosten C, Gunther S, Preiser W, et al. Identification of a novel coronavirus in patients with severe acute respiratory syndrome. *N Engl J Med* 2003;348:1967–1976.
- Ksiazek TG, Erdman D, Goldsmith CS, et al. A novel coronavirus associated with severe acute respiratory syndrome. *N Engl J Med* 2003;348:1953–1966.
- Peiris JS, Lai ST, Poon LL, et al. Coronavirus as a possible cause of severe acute respiratory syndrome. *Lancet* 2003;361:1319–1325.
- Christian MD, Poutanen SM, Loutfy MR, et al. Severe acute respiratory syndrome. *Clin Infect Dis* 2004;38:1420–1427.
- World Health Organization. Summary of probable SARS cases with onset of illness from 1 November 2002 to 31 July 2003. [http://www.who.int/csr/sars/country/table2004\\_04\\_21/en](http://www.who.int/csr/sars/country/table2004_04_21/en).
- Chinese SARS Molecular Epidemiology Consortium. Molecular evolution of the SARS coronavirus during the course of the SARS epidemic in China. *Science* 2004;303:1666–1669.
- Guan Y, Zheng BJ, He YQ, et al. Isolation and characterization of viruses related to the SARS coronavirus from animals in southern China. *Science* 2003;302:276–278.
- Lau SK, Woo PC, Li KS, et al. Severe acute respiratory syndrome coronavirus-like virus in Chinese horseshoe bats. *Proc Natl Acad Sci USA* 2005;102:14040–14045.
- Li W, Shi Z, Yu M, et al. Bats are natural reservoirs of SARS-like coronaviruses. *Science* 2005;310:676–679.
- Cheng VC, Lau SK, Woo PC, et al. Severe acute respiratory syndrome coronavirus as an agent of emerging and reemerging infection. *Clin Microbiol Rev* 2007;20:660–694.
- Ge XY, Li JL, Yang XL, et al. Isolation and characterization of a bat SARS-like coronavirus that uses the ACE2 receptor. *Nature* 2013;503:535–538.
- van der Hoek L, Pyrc K, Jebbink MF, et al. Identification of a new human coronavirus. *Nat Med* 2004;10:368–373.
- Fouchier RA, Hartwig NG, Bestebroer TM, et al. A previously undescribed coronavirus associated with respiratory disease in humans. *Proc Natl Acad Sci USA* 2004;101:6212–6216.
- Woo PC, Lau SK, Chu CM, et al. Characterization and complete genome sequence of a novel coronavirus, coronavirus HKU1, from patients with pneumonia. *J Virol* 2005;79:884–895.
- Talbot HK, Shepherd BE, Crowe JE Jr, et al. The pediatric burden of human coronaviruses evaluated for twenty years. *Pediatr Infect Dis J* 2009;28:682–687.
- Esper F, Weibel C, Ferguson D, et al. Coronavirus HKU1 infection in the United States. *Emerg Infect Dis* 2006;12:775–779.
- Zaki AM, van Boheemen S, Bestebroer TM, et al. Isolation of a novel coronavirus from a man with pneumonia in Saudi Arabia. *N Engl J Med* 2012;367:1814–1820.
- Assiri A, Al-Tawfiq JA, Al-Rabeeh AA, et al. Epidemiological, demographic, and clinical characteristics of 47 cases of Middle East respiratory syndrome coronavirus disease from Saudi Arabia: a descriptive study. *Lancet Infect Dis* 2013;13:752–761.
- Hu B, Ge X, Wang LF, Shi Z. Bat origin of human coronaviruses. *Virol J* 2015;12:221.
- World Health Organization. Middle East respiratory syndrome coronavirus (MERS-CoV). <http://www.who.int/emergencies/mers-cov/en/>.
- Vabret A, Dina J, Gouarin S, et al. Human (non-severe acute respiratory syndrome) coronavirus infections in hospitalised children in France. *J Paediatr Child Health* 2008;44:176–181.
- Dijkman R, Jebbink MF, El Drissi NB, et al. Human coronavirus NL63 and 229E seroconversion in children. *J Clin Microbiol* 2008;46:2368–2373.
- Severance EG, Bossis I, Dickerson FB, et al. Development of a nucleocapsid-based human coronavirus immunoassay and estimates of individuals exposed to coronavirus in a U.S. metropolitan population. *Clin Vaccine Immunol* 2008;15:1805–1810.
- Liang G, Chen Q, Xu J, et al. Laboratory diagnosis of four recent sporadic cases of community-acquired SARS, Guangdong Province, China. *Emerg Infect Dis* 2004;10:1774–1781.
- Lim PL, Kurup A, Gopalakrishna G, et al. Laboratory-acquired severe acute respiratory syndrome. *N Engl J Med* 2004;350:1740–1745.
- World Health Organization. SARS case in laboratory worker in Taiwan, China. <http://www.who.int/mediacentre/releases/2003/np26/en>.
- ProMED-mail. SARS—Worldwide (30): China, Cases. ProMED-mail. <http://www.promedmail.org> (archive number 20040703.1774).
- Wong T, Wallington T, McDonald LC, et al. Late recognition of SARS in nosocomial outbreak, Toronto. *Emerg Infect Dis* 2005;11:322–325.
- Alraddadi BM, Watson JT, Almarashi A, et al. Risk factors for primary Middle East respiratory syndrome coronavirus illness in humans, Saudi Arabia, 2014. *Emerg Infect Dis* 2016;22:49–55.
- Chowell G, Abdirizak F, Lee S, et al. Transmission characteristics of MERS and SARS in the healthcare setting: a comparative study. *BMC Med* 2015;13:210.
- Assiri A, McGeer A, Perl TM, et al. Hospital outbreak of Middle East respiratory syndrome coronavirus. *N Engl J Med* 2013;369:407–416.
- Memish ZA, Zumla AI, Al-Hakeem RF, et al. Family cluster of Middle East respiratory syndrome coronavirus infections. *N Engl J Med* 2013;368:2487–2494.
- World Health Organization. Middle East respiratory syndrome coronavirus (MERS-CoV)—Republic of Korea. <http://www.who.int/csr/don/21-july-2015-mers-korea/en/>.
- Goldmann DA. Transmission of viral respiratory infections in the home. *Pediatr Infect Dis J* 2000;19:S97–S102.
- Otter JA, Donskey C, Yezli S, et al. Transmission of SARS and MERS coronaviruses and influenza virus in healthcare settings: the possible role of dry surface contamination. *J Hosp Infect* 2016;92:235–250.
- Warnes SL, Little ZR, Keevil CW. Human coronavirus 229E remains infectious on common touch surface materials. *MBio* 2015;6(6):e1697–e1715.
- Lau JT, Lau M, Kim JH, et al. Probable secondary infections in households of SARS patients in Hong Kong. *Emerg Infect Dis* 2004;10:235–243.
- Loeb M, McGeer A, Henry B, et al. SARS among critical care nurses, Toronto. *Emerg Infect Dis* 2004;10:251–255.
- Scales DC, Green K, Chan AK, et al. Illness in intensive care staff after brief exposure to severe acute respiratory syndrome. *Emerg Infect Dis* 2003;9:1205–1210.
- Wong TW, Lee CK, Tam W, et al. Cluster of SARS among medical students exposed to single patient, Hong Kong. *Emerg Infect Dis* 2004;10:269–276.
- Yu IT, Li Y, Wong TW, et al. Evidence of airborne transmission of the severe acute respiratory syndrome virus. *N Engl J Med* 2004;350:1731–1739.
- Olsen SJ, Chang HL, Cheung TY, et al. Transmission of the severe acute respiratory syndrome on aircraft. *N Engl J Med* 2003;349:2416–2422.
- Chen YC, Huang LM, Chan CC, et al. SARS in hospital emergency room. *Emerg Infect Dis* 2004;10:782–788.
- Dowell SF, Simmerman JM, Erdman DD, et al. Severe acute respiratory syndrome coronavirus on hospital surfaces. *Clin Infect Dis* 2004;39:652–657.
- Wong SF, Chow KM, Leung TN, et al. Pregnancy and perinatal outcomes of women with severe acute respiratory syndrome. *Am J Obstet Gynecol* 2004;191:292–297.
- Shek CC, Ng PC, Fung GP, et al. Infants born to mothers with severe acute respiratory syndrome. *Pediatrics* 2003;112:e254.
- Bin SY, Heo JY, Song MS, et al. Environmental contamination and viral shedding in MERS patients during MERS-CoV outbreak in South Korea. *Clin Infect Dis* 2016;62:755–760.
- van Elden LJ, van Loon AM, van Alphen F, et al. Frequent detection of human coronaviruses in clinical specimens from patients with respiratory tract infection by use of a novel real-time reverse-transcriptase polymerase chain reaction. *J Infect Dis* 2004;189:652–657.
- Bradburne AF, Bynoe ML, Tyrrell DA. Effects of a “new” human respiratory virus in volunteers. *Br Med J* 1967;3:767–769.
- World Health Organization. Consensus document on the epidemiology of severe acute respiratory syndrome (SARS). <http://www.who.int/csr/sars/en/WHOconsensus.pdf>.
- Peiris JS, Chu CM, Cheng VC, et al. Clinical progression and viral load in a community outbreak of coronavirus-associated SARS pneumonia: a prospective study. *Lancet* 2003;361:1767–1772.
- Cheng PK, Wong DA, Tong LK, et al. Viral shedding patterns of coronavirus in patients with probable severe acute respiratory syndrome. *Lancet* 2004;363:1699–1700.
- Corman VM, Albarak AM, Omrani AS, et al. Viral shedding and antibody response in 37 patients with Middle East respiratory syndrome coronavirus infection. *Clin Infect Dis* 2016;62:477–483.
- Lessler J, Reich NG, Brookmeyer R, et al. Incubation periods of acute respiratory viral infections: a systematic review. *Lancet Infect Dis* 2009;9:291–300.
- Denison MR. Severe acute respiratory syndrome coronavirus pathogenesis, disease and vaccines: an update. *Pediatr Infect Dis J* 2004;23:S207–S214.
- Akerlund A, Greiff L, Andersson M, et al. Mucosal exudation of fibrinogen in coronavirus-induced common colds. *Acta Otolaryngol* 1993;113:642–648.
- Linden M, Greiff L, Andersson M, et al. Nasal cytokines in common cold and allergic rhinitis. *Clin Exp Allergy* 1995;25:166–172.
- Tyrrell DA, Cohen S, Schlarb JE. Signs and symptoms in common colds. *Epidemiol Infect* 1993;111:143–156.
- Callow KA, Parry HF, Sergeant M, et al. The time course of the immune response to experimental coronavirus infection of man. *Epidemiol Infect* 1990;105:435–446.
- Monto AS, Lim SK. The Tecumseh study of respiratory illness: VI. Frequency of and relationship between outbreaks of coronavirus infection. *J Infect Dis* 1974;129:271–276.
- Callow KA. Effect of specific humoral immunity and some non-specific factors on resistance of volunteers to respiratory coronavirus infection. *J Hyg (Lond)* 1985;95:173–189.
- Li W, Moore MJ, Vasilieva N, et al. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. *Nature* 2003;426(6965):450–454.
- Ng EK, Hui DS, Chan KC, et al. Quantitative analysis and prognostic implication of SARS coronavirus RNA in the plasma and serum of patients with severe acute respiratory syndrome. *Clin Chem* 2003;49:1976–1980.

73. Chan KH, Poon LL, Cheng VC, et al. Detection of SARS coronavirus in patients with suspected SARS. *Emerg Infect Dis* 2004;10:294–299.
74. Ding Y, He L, Zhang Q, et al. Organ distribution of severe acute respiratory syndrome (SARS) associated coronavirus (SARS-CoV) in SARS patients: implications for pathogenesis and virus transmission pathways. *J Pathol* 2004;203:622–630.
75. Farcas GA, Poutanen SM, Mazzulli T, et al. Fatal severe acute respiratory syndrome is associated with multiorgan involvement by coronavirus. *J Infect Dis* 2005;191:193–197.
76. Liu X, Shi Y, Li P, et al. Profile of antibodies to the nucleocapsid protein of the severe acute respiratory syndrome (SARS)-associated coronavirus in probable SARS patients. *Clin Diagn Lab Immunol* 2004;11:227–228.
77. Wong CK, Lam CW, Wu AK, et al. Plasma inflammatory cytokines and chemokines in severe acute respiratory syndrome. *Clin Exp Immunol* 2004;136:95–103.
78. Zhang Y, Li J, Zhan Y, et al. Analysis of serum cytokines in patients with severe acute respiratory syndrome. *Infect Immun* 2004;72:4410–4415.
79. Ng DL, Al HF, Keating MK, et al. Clinicopathologic, immunohistochemical, and ultrastructural findings of a fatal case of Middle East respiratory syndrome coronavirus infection in the United Arab Emirates, April 2014. *Am J Pathol* 2016;186:652–658.
80. Hocke AC, Becher A, Knepper J, et al. Emerging human Middle East respiratory syndrome coronavirus causes widespread infection and alveolar damage in human lungs. *Am J Respir Crit Care Med* 2013;188:882–886.
81. Hendley JO, Fishburne HB, Gwaltney JM Jr. Coronavirus infections in working adults: eight-year study with 229 E and OC 43. *Am Rev Respir Dis* 1972;105:805–811.
82. Bastien N, Robinson JL, Tse A, et al. Human coronavirus NL-63 infections in children: a 1-year study. *J Clin Microbiol* 2005;43:4567–4573.
83. Bastien N, Anderson K, Hart L, et al. Human coronavirus NL63 infection in Canada. *J Infect Dis* 2005;191:503–506.
84. Lau SK, Woo PC, Yip CC, et al. Coronavirus HKU1 and other coronavirus infections in Hong Kong. *J Clin Microbiol* 2006;44:2063–2071.
85. Makela MJ, Puhakka T, Ruuskanen O, et al. Viruses and bacteria in the etiology of the common cold. *J Clin Microbiol* 1998;36:539–542.
86. van der Zalm MM, van Ewijk BE, Wilbrink B, et al. Respiratory pathogens in children with and without respiratory symptoms. *J Pediatr* 2009;154:396–400.
87. Pappas DE, Hendley JO, Hayden FG, et al. Symptom profile of common colds in school-aged children. *Pediatr Infect Dis J* 2008;27:8–11.
88. Johnston SL, Pattemore PK, Sanderson G, et al. Community study of role of viral infections in exacerbations of asthma in 9–11 year old children. *BMJ* 1995;310:1225–1229.
89. Vabret A, Dina J, Gouarin S, et al. Detection of the new human coronavirus HKU1: a report of 6 cases. *Clin Infect Dis* 2006;42:634–639.
90. Falsey AR, Walsh EE, Hayden FG. Rhinovirus and coronavirus infection-associated hospitalizations among older adults. *J Infect Dis* 2002;185:1338–1341.
91. McIntosh K, Chao RK, Krause HE, et al. Coronavirus infection in acute lower respiratory tract disease of infants. *J Infect Dis* 1974;130:502–507.
92. Pene F, Merlat A, Vabret A, et al. Coronavirus 229E-related pneumonia in immunocompromised patients. *Clin Infect Dis* 2003;37:929–932.
93. Riski H, Hovi T. Coronavirus infections of man associated with diseases other than the common cold. *J Med Virol* 1980;6:259–265.
94. Arden KE, Nissen MD, Sloots TP, et al. New human coronavirus, HCoV-NL63, associated with severe lower respiratory tract disease in Australia. *J Med Virol* 2005;75:455–462.
95. Esper F, Weibel C, Ferguson D, et al. Evidence of a novel human coronavirus that is associated with respiratory tract disease in infants and young children. *J Infect Dis* 2005;191:492–498.
96. Chiu SS, Chan KH, Chu KW, et al. Human coronavirus NL63 infection and other coronavirus infections in children hospitalized with acute respiratory disease in Hong Kong, China. *Clin Infect Dis* 2005;40:1721–1729.
97. Ebihara T, Endo R, Ma X, et al. Detection of human coronavirus NL63 in young children with bronchiolitis. *J Med Virol* 2005;75:463–465.
98. Moes E, Vijgen L, Keyaerts E, et al. A novel pancoronavirus RT-PCR assay: frequent detection of human coronavirus NL63 in children hospitalized with respiratory tract infections in Belgium. *BMC Infect Dis* 2005;5:6.
99. Sung JY, Lee HJ, Eun BW, et al. Role of human coronavirus NL63 in hospitalized children with croup. *Pediatr Infect Dis J* 2010;29:822–826.
100. van der Hoek L, Sure K, Ithor G, et al. Croup is associated with the novel coronavirus NL63. *PLoS Med* 2005;2:e240.
101. Chang LY, Chiang BL, Kao CL, et al. Lack of association between infection with a novel human coronavirus (HCoV), HCoV-NH, and Kawasaki disease in Taiwan. *J Infect Dis* 2006;193:283–286.
102. Esper F, Shapiro ED, Weibel C, et al. Association between a novel human coronavirus and Kawasaki disease. *J Infect Dis* 2005;191:499–502.
103. Esper F, Ou Z, Huang YT. Human coronaviruses are uncommon in patients with gastrointestinal illness. *J Clin Virol* 2010;48:131–133.
104. Tsui PT, Kwok ML, Yuen H, et al. Severe acute respiratory syndrome: clinical outcome and prognostic correlates. *Emerg Infect Dis* 2003;9:1064–1069.
105. Fowler RA, Lapinsky SE, Hallett D, et al. Critically ill patients with severe acute respiratory syndrome. *JAMA* 2003;290:367–373.
106. Lew TW, Kwek TK, Tai D, et al. Acute respiratory distress syndrome in critically ill patients with severe acute respiratory syndrome. *JAMA* 2003;290:374–380.
107. Donnelly CA, Ghani AC, Leung GM, et al. Epidemiological determinants of spread of causal agent of severe acute respiratory syndrome in Hong Kong. *Lancet* 2003;361:1761–1766.
108. Booth CM, Matukas LM, Tomlinson GA, et al. Clinical features and short-term outcomes of 144 patients with SARS in the greater Toronto area. *JAMA* 2003;289:2801–2809.
109. Wang JT, Sheng WH, Fang CT, et al. Clinical manifestations, laboratory findings, and treatment outcomes of SARS patients. *Emerg Infect Dis* 2004;10:818–824.
110. Wong KT, Antonio GE, Hui DS, et al. Severe acute respiratory syndrome: radiographic appearances and pattern of progression in 138 patients. *Radiology* 2003;228:401–406.
111. Muller NL, Ooi GC, Khong PL, et al. Severe acute respiratory syndrome: radiographic and CT findings. *AJR Am J Roentgenol* 2003;181:3–8.
112. Wong KT, Antonio GE, Hui DS, et al. Thin-section CT of severe acute respiratory syndrome: evaluation of 73 patients exposed to or with the disease. *Radiology* 2003;228:395–400.
113. Bitnun A, Allen U, Heurter H, et al. Children hospitalized with severe acute respiratory syndrome-related illness in Toronto. *Pediatrics* 2003;112:e261.
114. Hon KL, Leung CW, Cheng WT, et al. Clinical presentations and outcome of severe acute respiratory syndrome in children. *Lancet* 2003;361:1701–1703.
115. Yang GG, Lin SZ, Liao KW, et al. SARS-associated coronavirus infection in teenagers. *Emerg Infect Dis* 2004;10:382–383.
116. Chiu WK, Cheung PC, Ng KL, et al. Severe acute respiratory syndrome in children: experience in a regional hospital in Hong Kong. *Pediatr Crit Care Med* 2003;4:279–283.
117. Ng PC, Leung CW, Chiu WK, et al. SARS in newborns and children. *Biol Neonate* 2004;85:293–298.
118. Arabi YM, Arif AA, Balkhy HH, et al. Clinical course and outcomes of critically ill patients with Middle East respiratory syndrome coronavirus infection. *Ann Intern Med* 2014;160:389–397.
119. Poon LL, Chan KH, Wong OK, et al. Early diagnosis of SARS coronavirus infection by real time RT-PCR. *J Clin Virol* 2003;28:233–238.
120. Chan PK, Ng KC, Chan RC, et al. Immunofluorescence assay for serologic diagnosis of SARS. *Emerg Infect Dis* 2004;10:530–532.
121. Alshukairi AN, Khalid I, Ahmed WA, et al. Antibody response and disease severity in healthcare worker MERS survivors. *Emerg Infect Dis* 2016;22(6):doi:10.3201/eid2206.160010.
122. Tang P, Louie M, Richardson SE, et al. Interpretation of diagnostic laboratory tests for severe acute respiratory syndrome: the Toronto experience. *Can Med Assoc J* 2004;170:47–54.
123. Grant PR, Garson JA, Tedder RS, et al. Detection of SARS coronavirus in plasma by real-time RT-PCR. *N Engl J Med* 2003;349:2468–2469.
124. Centers for Disease Control and Prevention. CDC laboratory testing for Middle East respiratory syndrome coronavirus (MERS-CoV). <http://www.cdc.gov/coronavirus/mers/lab/lab-testing.html>.
125. Centers for Disease Control and Prevention. Public health guidance for community-level preparedness and response to severe acute respiratory syndrome (SARS), version 2/3. Appendix F8: guidelines for laboratory diagnosis of SARS-CoV infection. Supplement F: laboratory guidance. <http://www.cdc.gov/sars/guidance/f-lab/app8.html>.
126. Chan KS, Lai ST, Chu CM, et al. Treatment of severe acute respiratory syndrome with lopinavir/ritonavir: a multicentre retrospective matched cohort study. *Hong Kong Med J* 2003;9:399–406.
127. Loutfy MR, Blatt LM, Siminovitch KA, et al. Interferon alfacon-1 plus corticosteroids in severe acute respiratory syndrome: a preliminary study. *JAMA* 2003;290:3222–3228.
128. Wong VW, Dai D, Wu AK, et al. Treatment of severe acute respiratory syndrome with convalescent plasma. *Hong Kong Med J* 2003;9:199–201.
129. Zhao Z, Zhang F, Xu M, et al. Description and clinical treatment of an early outbreak of severe acute respiratory syndrome (SARS) in Guangzhou, PR China. *J Med Microbiol* 2003;52:715–720.
130. Barnard DL, Hubbard VD, Burton J, et al. Inhibition of severe acute respiratory syndrome-associated coronavirus (SARSCoV) by calpain inhibitors and beta-D-N4-hydroxycytidine. *Antivir Chem Chemother* 2004;15:15–22.
131. Chu CM, Cheng VC, Hung IF, et al. Role of lopinavir/ritonavir in the treatment of SARS: initial virological and clinical findings. *Thorax* 2004;59:252–256.
132. Cinatl J Jr, Michaelis M, Scholz M, et al. Role of interferons in the treatment of severe acute respiratory syndrome. *Expert Opin Biol Ther* 2004;4:827–836.
133. Haagmans BL, Kuiken T, Martina BE, et al. Pegylated interferon-alpha protects type I pneumocytes against SARS coronavirus infection in macaques. *Nat Med* 2004;10:290–293.
134. Hensley LE, Fritz LE, Jahrling PB, et al. Interferon-beta 1a and SARS coronavirus replication. *Emerg Infect Dis* 2004;10:317–319.
135. Stroher U, DiCaro A, Li Y, et al. Severe acute respiratory syndrome-related coronavirus is inhibited by interferon-alpha. *J Infect Dis* 2004;189:1164–1167.
136. Tan EL, Ooi EE, Lin CY, et al. Inhibition of SARS coronavirus infection in vitro with clinically approved antiviral drugs. *Emerg Infect Dis* 2004;10:581–586.
137. Yamamoto N, Yang R, Yoshinaka Y, et al. HIV protease inhibitor nelfinavir inhibits replication of SARS-associated coronavirus. *Biochem Biophys Res Commun* 2004;318:719–725.
138. Zheng B, He ML, Wong KL, et al. Potent inhibition of SARS-associated coronavirus (SCoV) infection and replication by type I interferons (IFN-alpha/beta) but not by type II interferon (IFN-gamma). *J Interferon Cytokine Res* 2004;24:388–390.
139. Cinatl J, Morgenstern B, Bauer G, et al. Glycyrrhizin, an active component of liquorice roots, and replication of SARS-associated coronavirus. *Lancet* 2003;361:2045–2046.
140. Zhaori G. Antiviral treatment of SARS: can we draw any conclusions? *Can Med Assoc J* 2003;169:1165–1166.

141. Groneberg DA, Poutanen SM, Low DE, et al. Treatment and vaccines for severe acute respiratory syndrome. *Lancet Infect Dis* 2005;5:147–155.
142. Public Health England. Treatment of MERS-CoV: information for clinicians. Clinical decision-making support for treatment of MERS-CoV patients. September 2015. [https://www.gov.uk/government/uploads/system/uploads/attachment\\_data/file/459835/merscov\\_for\\_clinicians\\_sept2015.pdf](https://www.gov.uk/government/uploads/system/uploads/attachment_data/file/459835/merscov_for_clinicians_sept2015.pdf).
143. Roberts L, Smith W, Jorm L, et al. Effect of infection control measures on the frequency of upper respiratory infection in child care: a randomized, controlled trial. *Pediatrics* 2000;105:738–742.
144. Apisarnthanarak A, Apisarnthanarak P, Cheevakumjorn B, et al. Implementation of an infection control bundle in a school to reduce transmission of influenza-like illness during the novel influenza A 2009 H1N1 pandemic. *Infect Control Hosp Epidemiol* 2010;31:310–311.
145. Turner RB, Felton A, Kosak K, et al. Prevention of experimental coronavirus colds with intranasal alpha-2b interferon. *J Infect Dis* 1986;154:443–447.
146. Higgins PG, Phillpotts RJ, Scott GM, et al. Intranasal interferon as protection against experimental respiratory coronavirus infection in volunteers. *Antimicrob Agents Chemother* 1983;24:713–715.
147. McElhaney JE, Goel V, Toane B, et al. Efficacy of COLD-fX in the prevention of respiratory symptoms in community-dwelling adults: a randomized, double-blinded, placebo controlled trial. *J Altern Complement Med* 2006;12:153–157.
148. Predy GN, Goel V, Lovlin R, et al. Efficacy of an extract of North American ginseng containing poly-furanosyl-pyranosyl-saccharides for preventing upper respiratory tract infections: a randomized controlled trial. *Can Med Assoc J* 2005;173:1043–1048.
149. Wang M, Guilbert LJ, Ling L, et al. Immunomodulating activity of CVT-E002, a proprietary extract from North American ginseng (*Panax quinquefolium*). *J Pharm Pharmacol* 2001;53:1515–1523.
150. Wang M, Guilbert LJ, Li J, et al. A proprietary extract from North American ginseng (*Panax quinquefolium*) enhances IL-2 and IFN-gamma productions in murine spleen cells induced by Con-A. *Int Immunopharmacol* 2004;4:311–315.
151. Siegel JD, Rhinehart E, Jackson M, et al. Centers for Disease Control and Prevention: 2007 guideline for isolation precautions: preventing transmission of infectious agents in healthcare settings. <http://www.cdc.gov/hicpac/pdf/isolation/Isolation2007.pdf>.
152. Centers for Disease Control and Prevention. Interim infection prevention and control recommendations for hospitalized patients with Middle East respiratory syndrome coronavirus (MERS-CoV). <https://www.cdc.gov/coronavirus/mers/infection-prevention-control.html>.
153. Rabenau HF, Cinatl J, Morgenstern B, et al. Stability and inactivation of SARS coronavirus. *Med Microbiol Immunol* 2005;194:1–6.
154. Roper RL, Rehm KE. SARS vaccines: where are we? *Expert Rev Vaccines* 2009;8:887–898.
155. Stadler K, Masignani V, Eickmann M, et al. SARS—beginning to understand a new virus. *Nat Rev Microbiol* 2003;1:209–218.
156. Omrani AS, Al-Tawfiq JA, Memish ZA. Middle East respiratory syndrome coronavirus (MERS-CoV): animal to human interaction. *Pathog Glob Health* 2015;109:354–362.
157. Wang L, Shi W, Joyce MG, et al. Evaluation of candidate vaccine approaches for MERS-CoV. *Nat Commun* 2015;6:7712.